## What is 'radiation quality'?

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## radiation quality?

Try the web:

radiation quality,

a descriptive specification of the **penetrating nature** of an x-ray beam. It is influenced by <a href="kilovoltage">kilovoltage</a> and **filtration**: a higher kilovoltage produces more penetration, and filtration removes selected wavelengths and "hardens" the beam. medical-dictionary.thefreedictionary.com

The ability of a beam of x-rays to allow the production of **diagnostically useful radiographs**.

Usually measured in **half-value layers** of aluminum and controlled by the kilovolt peak.

Mosby's Dental Dictionary

The spectrum of radiant energy produced by a given radiation source with respect to its **penetration** or its **suitability** for a specific application.

McGraw-Hill Science & Technology Dictionary

#### Such 'definitions' depend on the application of interest ??

Mostly regarding radiography/imaging rather than biology/health effects

Try: International Commission on Radiological Protection (ICRP)
International Commission on Radiation Units and Measurements (ICRU)

ICRP Publication 92 (2003): Relative Biological Effectiveness (RBE), Quality Factor (Q), and Radiation Weighting Factor ( $w_R$ )

ICRP Publication 103 (2007): The 2007 Recommendations of the ICRP.

ICRU Report 16 (1970): Linear Energy Transfer.

ICRU Report 36 (1983): Microdosimetry.

ICRU Report 40 (1986): The Quality Factor in Radiation Protection.

ICRU Report ICRU 60 & 85 (1998 & 2011). Fundamental Quantities and Units for Ionizing Radiation.

ICRP Publication 60 (1991): 1990 Recommendations of the ICRP

"The probability of stochastic effects is found to depend, not only on the absorbed dose, but also on the type and energy of the radiation causing the dose. This is taken into account by weighting the absorbed dose by a factor related to the quality of the radiation."

But, what is "quality of the radiation"?

When x rays and y rays were the only types of ionizing radiation available to the therapist, the term "quality" was used to describe the penetrating power of the radiation. Quality was usually expressed in terms of the half-value layer in copper or aluminium (i.e., the thickness of material required to reduce the intensity to half). With the extension of radio-therapy and radiobiology to other types of radiation it was realized that the biological effect per unit absorbed dose depended on the radiation used. The term "quality" became a description of the radiation as it affects the biological

(Bewley 1973)

response;

From "Radiation Quality and its Influence on Biological Response"

"The pioneering experiments by Zirkle (1935) and a multitude of succeeding studies have established that the biological effectiveness of ionizing radiation depends not only on the amount of energy absorbed but also on the spatial distribution of energy deposition. Since the energy is imparted in or near the tracks of charged particles, it has been considered convenient to express the heterogeneity of energy deposition in terms of the linear density of energy loss in these tracks. The term linear energy transfer (LET) has been coined by Zirkle et al. (1952) for this purpose. Using this concept one may express radiation quality as a distribution of dose in LET, specifying the fraction of the dose deposited in each LET interval."

From "Specification of Radiation Quality" (Rossi 1959)

• We now know, of course, that LET is far from adequate to specify radiation quality.

( LET = Linear Energy Transfer )

## To describe radiation effects or mechanisms, need physical specification of:

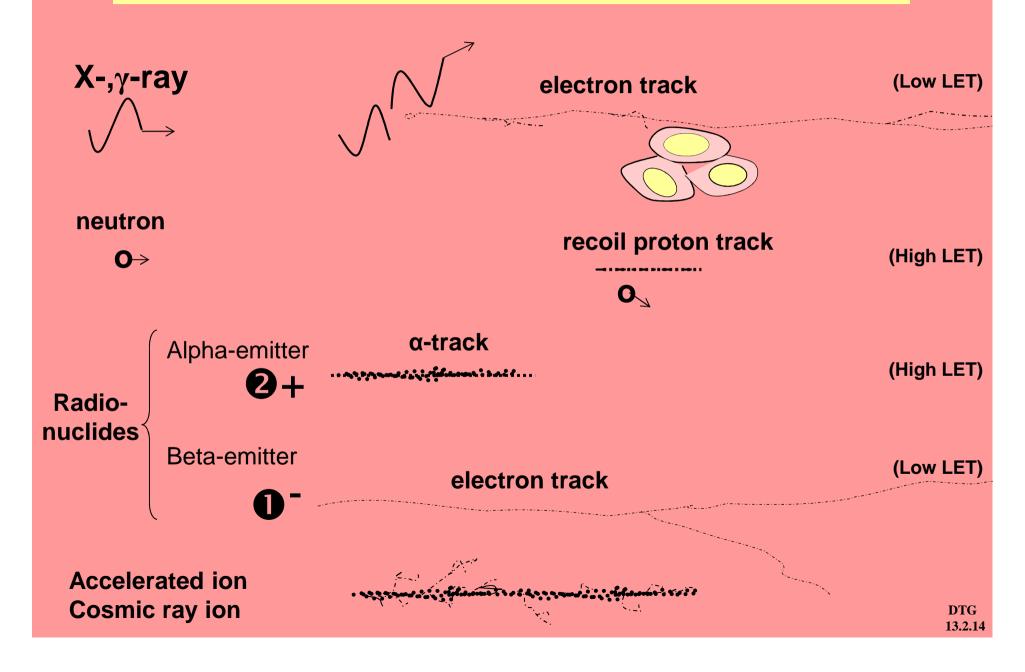
- 1. Measure of <u>quantity</u> of radiation: -- Use absorbed dose, or fluence
- 2. Variations of dose on scale of interest e.g. isodose plots; tissue compartments; doses on more microscopic scale from internal radionuclides;
- 3. <u>Time</u> course of delivery: e.g. Dose rate, fractionation or fluence rate, etc
- 4. Specification of 'quality' of the radiation:

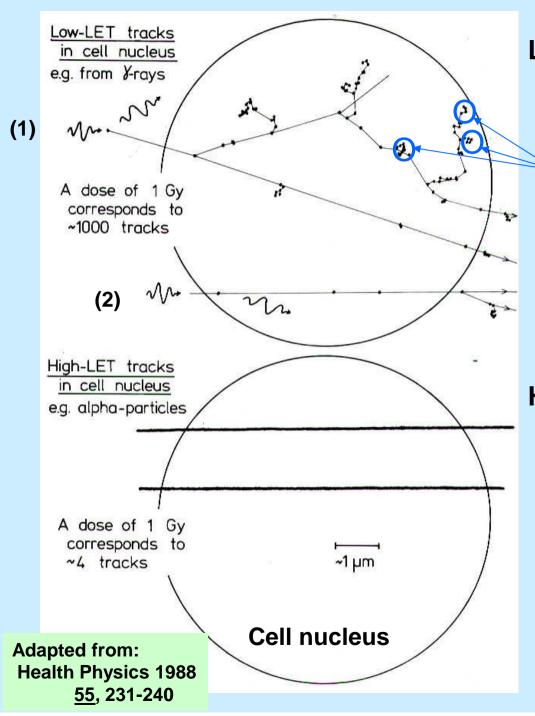
  LET as 1<sup>st</sup> approximation.

  Better options?

Of course, effects depend also on the particular biological system itself and its environment.

## The insult to DNA, cells and tissue from ionizing radiation is always in the form of structured tracks from charged particles





#### **Low-LET radiation:**

Sparsely ionizing on average,
but ~ 1/4 of energy deposited via
denser clusters of ionizations
from low-energy secondary
electrons (on scale of nanometres)
(Magnified in diagram)

Very low dose from a single track (~ 0.001 Gy to cell nucleus)

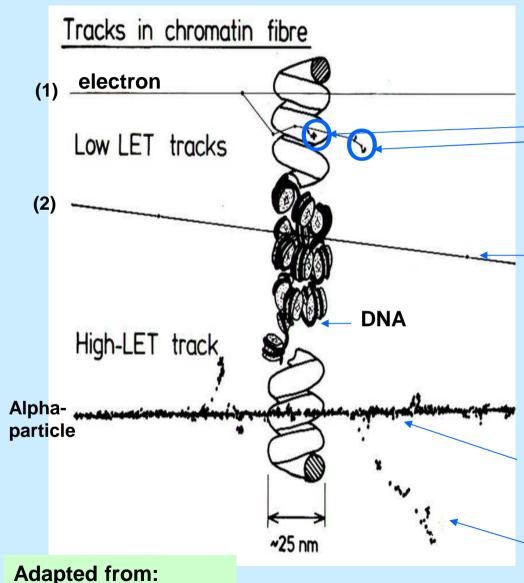
#### **High-LET radiation:**

Densely ionizing on average (especially for low-velocity ions, natural alpha-particles, etc)

High dose from a single track ( $\sim 0.2 - 0.5$  Gy from single  $\alpha$ -track)

LET = <u>Linear Energy Transfer</u>

#### All radiation tracks are highly structured on the scale of DNA



Clustered ionizations from low-energy electron

Single ionization

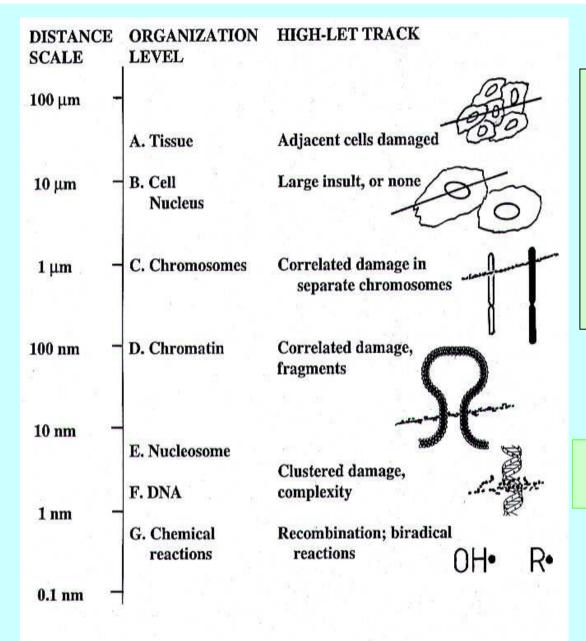
**Opposing trends:** Alpha-particle has

- -- low probability of hitting DNA (few tracks per Gy)
- -- high probability of damage when it does hit.

Dense ionization clustering along path of alpha particle

**Delta-ray electron** 

Adapted from: Health Physics 1988 55, 231-240



Radiation track structure is important at all levels of organisation, from molecules to tissue, from sub-nanometres to 100s of micrometres

The DNA level (nanometres) is particularly important.

High-LET and low-LET radiations are different at <u>all</u> these levels. Which level(s) dominate the biological effectiveness?

#### **Absorbed dose** of ionizing radiation is:

- the amount of energy imparted per unit mass of tissue.
- measured in units of joules per kilogramme, given the special name gray.

$$1 Gy = 1 J/kg$$

**ICRU Definition:** 

4.2.5 Absorbed Dose

The **absorbed dose**, D, is the quotient of  $d\bar{\epsilon}$  by dm, where  $d\bar{\epsilon}$  is the mean energy imparted to matter of mass dm, thus

$$D = \frac{\mathrm{d}\overline{\epsilon}}{\mathrm{d}m} \,.$$

Unit: J kg<sup>-1</sup>

The special name for the unit of absorbed dose is gray (Gy).

Usually applied as an <u>average</u> in a <u>macroscopic</u> mass (volume) of tissue

Ignores microscopic variations and stochastics

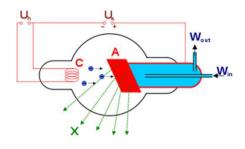
# Radiation Quality is defined by the <u>fluence spectrum</u> of radiation particles at the locations of interest in the target material. (biological system)

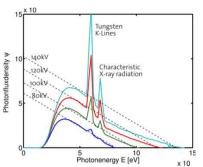
 Depends on characteristics of the radiation source and the intervening material

#### Fluence spectrum:

- specifies the <u>relative</u> numbers of <u>particles according to type and energy</u>
- includes:
- charged particles ---- of particular importance for most biological effects (e.g. electrons, protons, alpha-particles, heavier ions)
- (neutral particles also, such as X- & γ-ray photons and neutrons)

### Radiation source characteristics



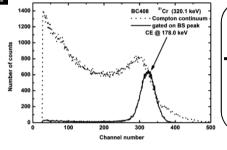


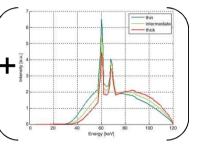
**Intervening material** 



RADIATION QUALITY

## Fluence spectrum of charged and neutral particles (particle types and energies)

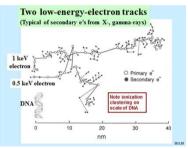


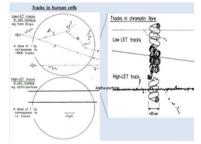


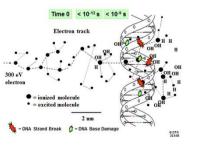
**Track structures** 

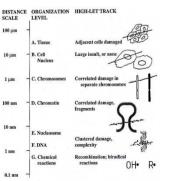


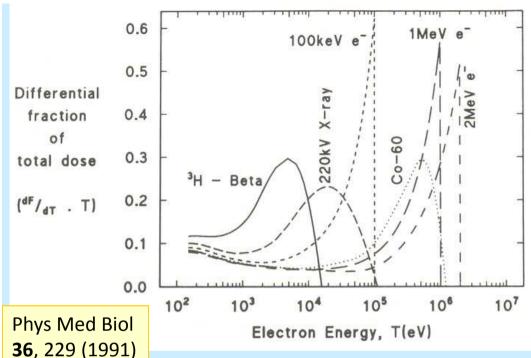
## Biological damage and health effects











#### Differences in quality between some **low-LET radiations**

Low-energy electrons are an important component for dose deposition by all **Iow-LET radiations (X-, y-rays, beta-emitters)** 

Such differences in radiation quality can be significant for biological effects

#### **COMPARING LOW-LET RADIATIONS:**

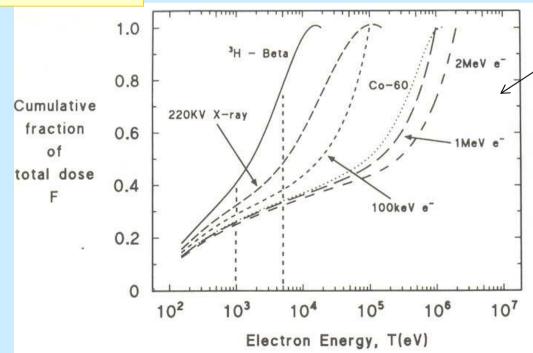
**Dose fraction deposited by electrons** of energies 0.1 to 5 keV

Tritium β	77 %	42%
220 kV X-rays	48 %	33%
Co γ-rays	34 %	27%

**NOTE:** Low energy electrons are more efficient at producing:

- DNA double-strand breaks (DSB)
- a higher proportion of complex DSB (and other clustered damage)
- a wide variety of biological effects in cells (mutations, chromosome aberrations, malignant transformation, killing, etc)

1.0



#### 5.3 MeV alpha particle

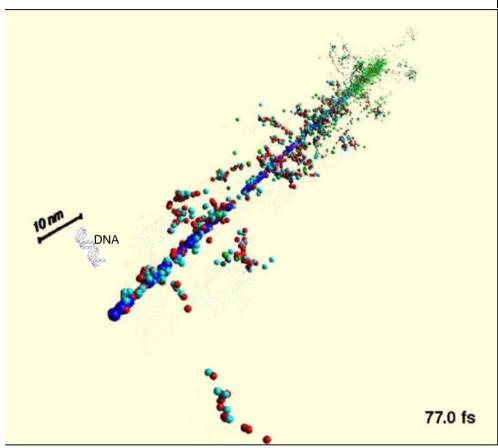
1.3 MeV/u

LET ~ 88 keV/µm

#### 200 MeV Oxygen ion

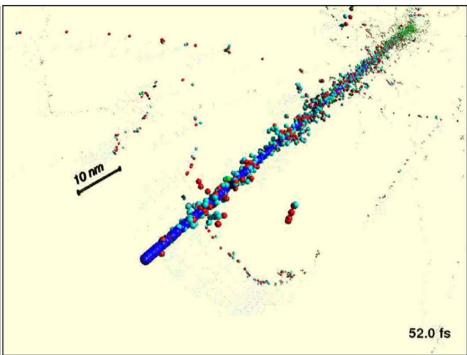
12.5 MeV/u

LET  $\sim$  250 keV/ $\mu$ m



By courtesy of Herwig Paretzke, Werner Friedland and Maximillian Kreipl

Track simulation methods in: Kreipl et al, Radiat Environm Biophys 48, 349-359 (2009).

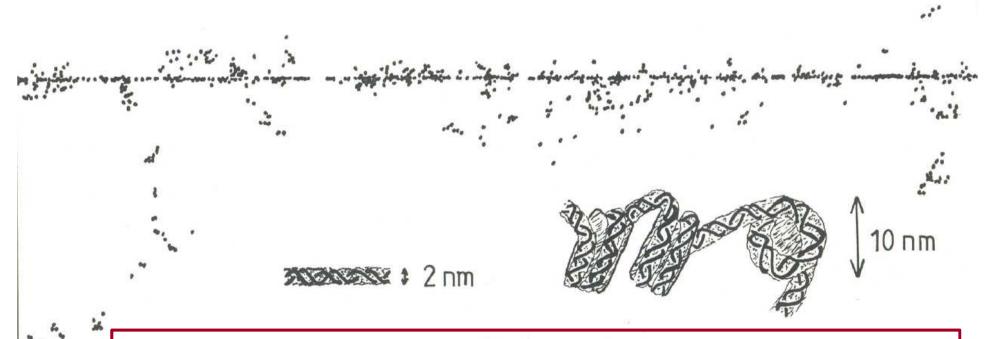


NOTE: Each symbol represents a <a href="point">point</a> interaction.

These diagrams use finite spheres to provide a perspective of distance.

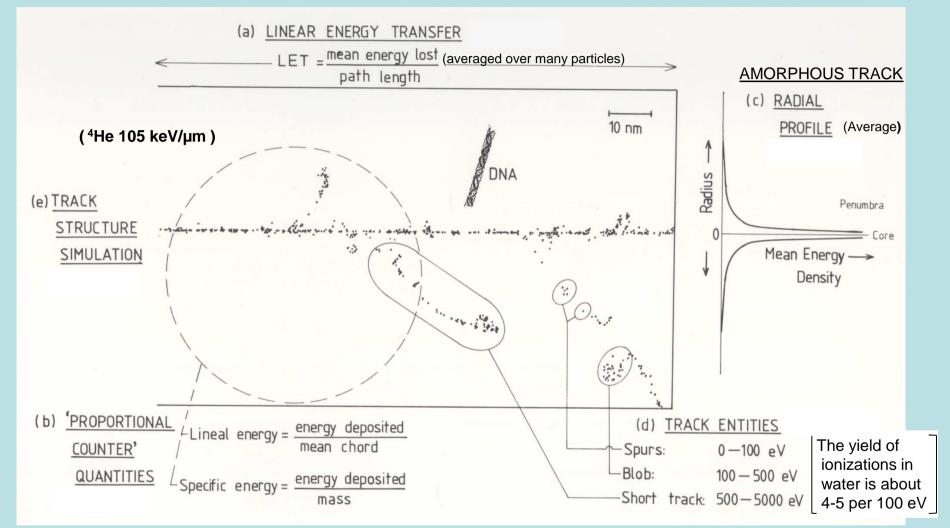
Shown are frozen sample screens from live simulations run for visual appreciation of track structure.

## A short segment of a 4 MeV <sup>4</sup>He (alpha-particle) track (105 keV/µm)



Need descriptors/parameters to relate physical 'radiation quality' to biological effectiveness

#### **Descriptions of radiation quality:**

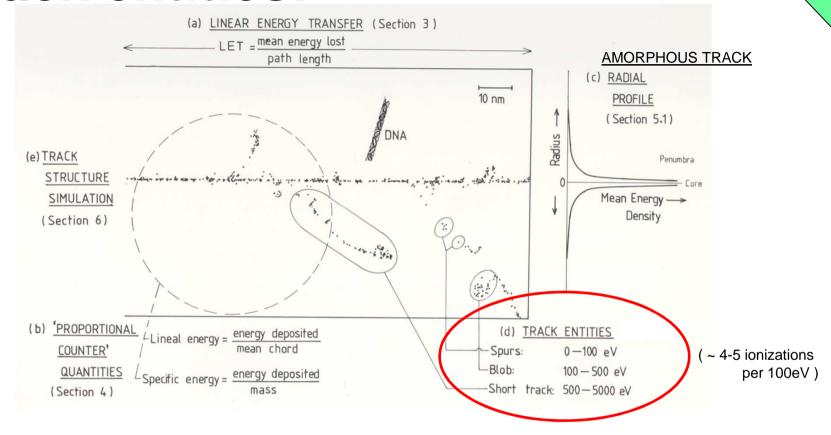


[ Adapted from Int J Radiat Biol <u>56</u>, 623 (1989)]

No single description is adequate or sufficient

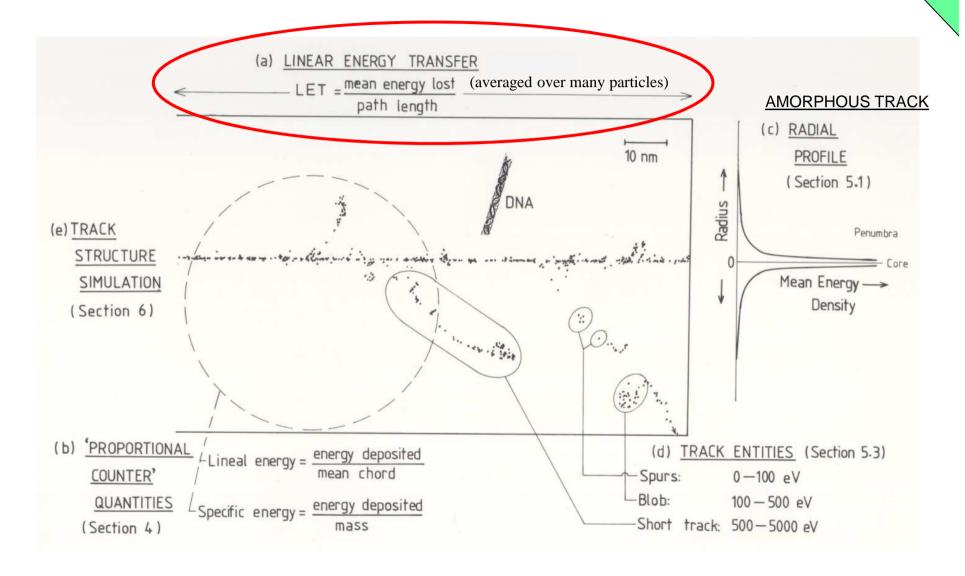
Track simulations give ~ complete description, but info must be distilled/reduced

### **Track entities:**



- Developed and used by radiation chemists (Mozumder & Magee1966: Radiat Res 28, 203-214)
- Little application in radiation biology (e.g. Ward 1981: Radiat Res 86, 185-195)
- No application in radiation protection or medicine.

## **Linear Energy Transfer (LET):**



### **Consider LET**

- Describes energy transfer (loss) <u>along</u> the path of the particle (Averaged over many particles of same Z, E)
- BUT gives NO information on:
  - -- Fluctuations in energy loss (stochastics)
  - -- Lateral spread of the track
- LET depends on particle charge (Z) and velocity (V)

Hence

Particles of same LET can have grossly different track structures

Linear electronic Stopping Power

Note: An alternative is to use  $z^2/\beta^2$  instead of LET, but generally similar limitations.

The linear energy transfer or restricted linear electronic stopping power,  $L_{\Delta}$ , of a material, for charged particles, is the quotient of  $dE_{\Delta}$  by dl, where  $dE_{\Delta}$  is the energy lost by a charged particle due to electronic collisions in traversing a distance dl, minus the sum of the kinetic energies of all the electrons released with kinetic energies in excess of  $\Delta$ , thus

$$L_{\Delta}=rac{\mathrm{d}E_{\Delta}}{\mathrm{d}l}\,.$$

Unit: J m<sup>-1</sup>

The cut-off value ( $\Delta$ ) is usually in electron-volts (eV)

Most commonly used are:

 $L_{\infty}$  i.e. no cut-off (unrestricted), simply written as L

L<sub>100</sub> i.e. 100 eV cut-off, includes only electrons of range ~nms (ie very local)

Hence, can define mean LET of a radiation field as:

Track-average LET:

$$\overline{L}_{\rm T} = \int_0^\infty Lt(L) \ dL / \int_0^\infty t(L) \ dL$$

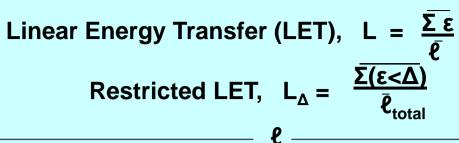
Use if effect of interest is  $\approx$  proportional to L.

**Dose average LET:** 

$$\overline{L}_{\rm D} = \int_0^\infty L^2 t(L) \ dL / \int_0^\infty L t(L) \ dL$$

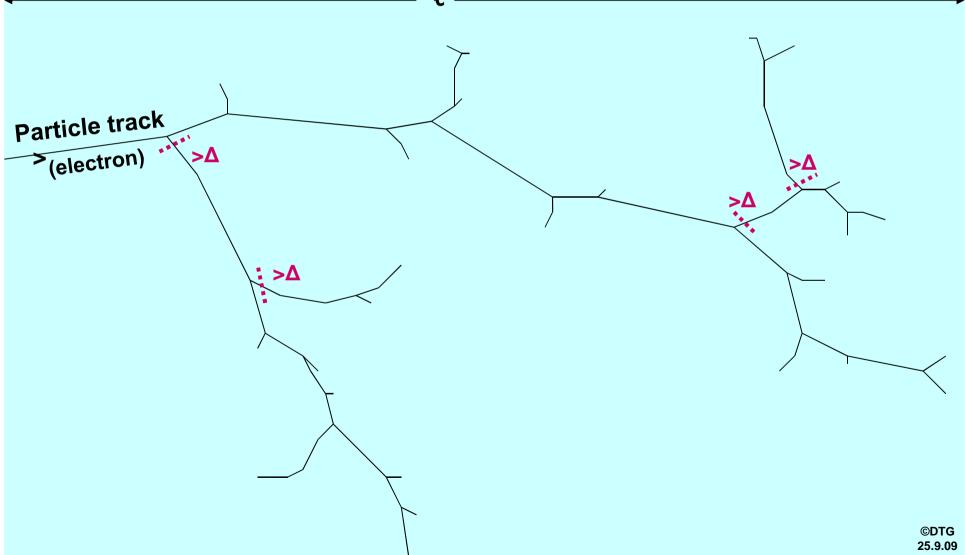
Use if effect of interest is  $\approx$  proportional to L<sup>2</sup>.

where t(L) is the frequency distribution of L in the field



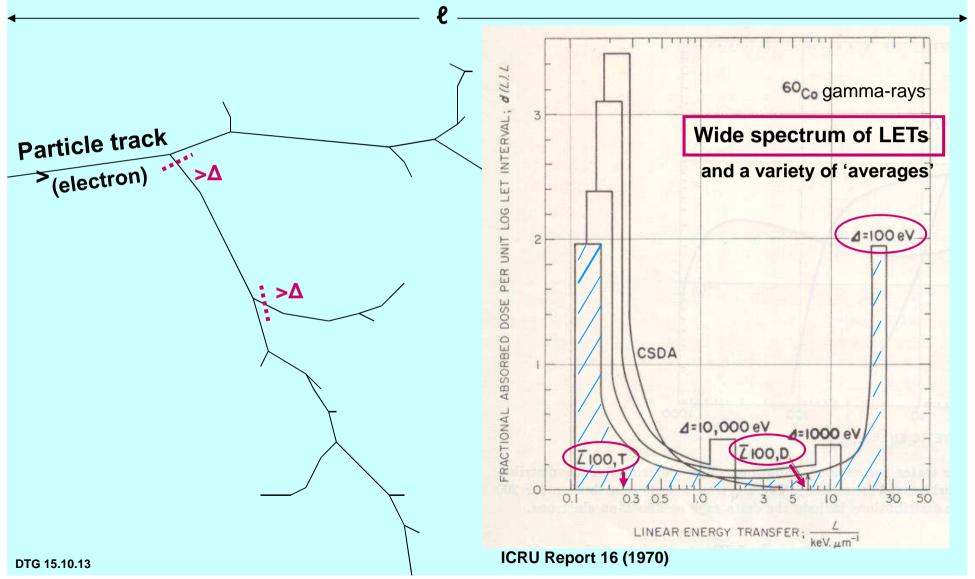
(Averaged over many tracks of this energy)

If ε>Δ Cut off and treat as separate track



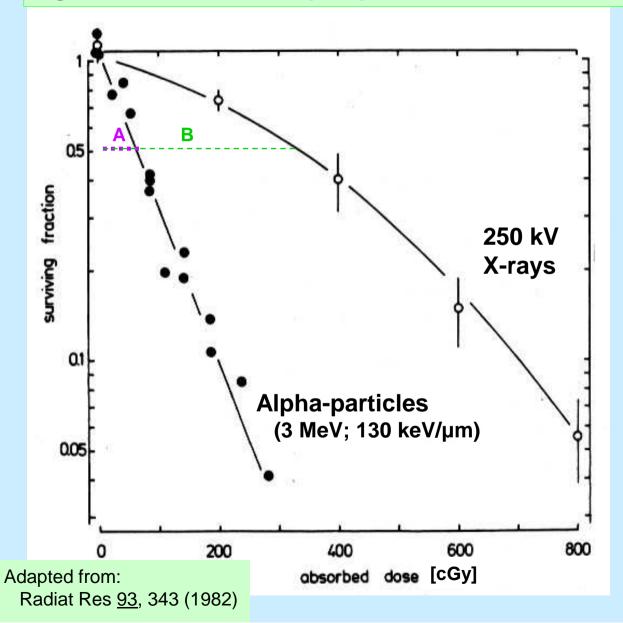
Linear Energy Transfer (LET),  $L = \frac{\overline{\Sigma \epsilon}}{\ell}$ Restricted LET,  $L_{\Delta} = \frac{\overline{\Sigma(\epsilon < \Delta)}}{\ell_{total}}$  (Averaged over many tracks of this energy)

If ε>Δ Cut off and treat as separate track



#### **Ionizing radiations can kill cells:**

#### e.g. Cell survival after alpha-particle irradiation compared to X-rays (in V79 cells)



For 50% survival of these cells the RBE\* of alpha-particles relative to X-rays is

 $\frac{\text{Dose B}}{\text{Dose A}} \sim 5$ 

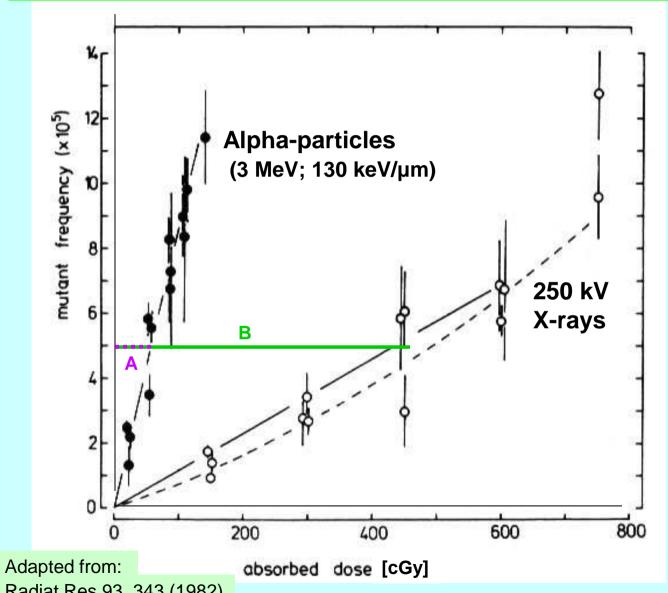
This RBE is dose-dependent

-- larger at lower doses

\* RBE = Relative Biological
Effectiveness
= Ratio of doses for
identical level of
biological effect

#### **Ionizing radiations mutate genes in cells:**

e.g. hprt mutation-induction by alpha-particles compared to X-rays (in V79 cells)



In general, biological effectiveness depends on:

- --- radiation quality
- --- dose
- --- dose-rate
- --- biological system

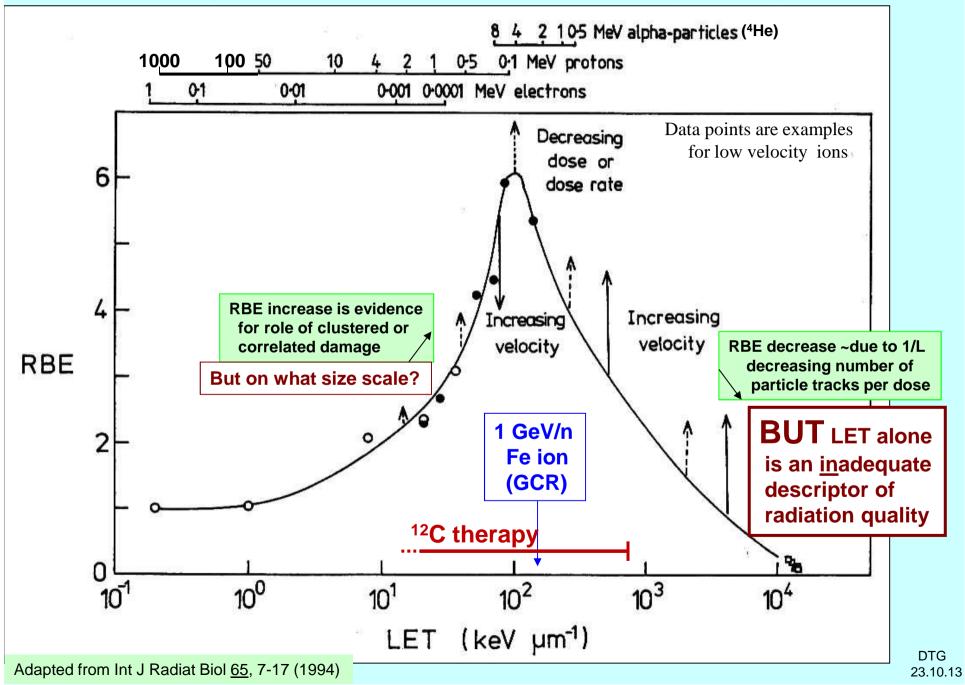
#### Here:

**Relative Biological** Effectiveness (RBE) of alpha-particles in this system is

> **Dose B** ~ 8 **Dose A**

Radiat Res <u>93</u>, 343 (1982)





#### Some applications of LET

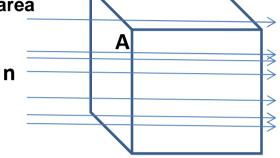
For fluence F of particles of LET L, the absorbed dose D is

D = kFL where F = n/A = number of particles/unit area

= 0.16FL for A ( $\mu$ m), D (Gy), L ( $keV/\mu$ m

Organise radiation-quality data

e.g. RBE versus LET plots



#### **Basic studies:**

• Early analyses of radiation action for cell killing, mutation, aberrations, etc e.g. Brustad (1962), Howard-Flanders (1958), Barendsen (1966), Goodhead (1980)

#### Radiation protection:

 Quality factor, Q(L), to convert absorbed dose to dose equivalent, in current operational radiation protection (monitoring)

#### Radiotherapy:

• General indicator of increasing effectiveness & decreasing OER (Oxygen Enhancement Ratio)

 RBE model for "biological dose" for application in treatment planning for heavy ion RT at HIMAC (Kanai et al RR, Radiat Res 147, 78-85 (1997))

Based on linear-quadratic survival dose-response, with parameters empirically dependent on LET.

#### In operational radiation protection:

## Q(L) relationship is used to calculate the operational dose equivalent used in monitoring

$$H = k \int Q(L) \left| L \frac{\partial \Phi(L)}{dL} \right| dL$$

Q weights absorbed dose (Gy) to obtain dose equivalent (Sv)

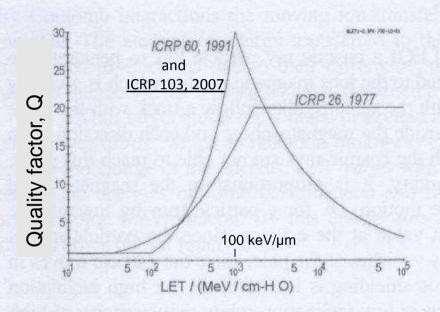


Figure 1.8 Quality factor, Q, as function of LET as defined by:

(ICRP 60, 1991 and ICRP 26, 1977)

Based on ICRP committee judgements from experimental/theoretical considerations, because ~no epidemiological data are available for most high-LET radiations.

yields the equivalent dose deposited at the given point with Q(L) representing the quality factor dependence on LET as in Figure 1.8 and k being an appropriate unit conversion factor.

Reliance on LET as the sole radiation-quality parameter is a notable limitation

--- All other aspects of track structure are ignored

Note: For most radiation protection ICRP-defined radiation weighting factors,  $w_R$ , are used to convert absorbed dose to equivalent dose (ICRP 103 (2007)).

#### ICRP-prescribed values of radiation weighting factor

NOTE: ~ all based on experimental/theoretical info, because ~no epi

Radiation type and energy range	Prescribed w <sub>R</sub>	
	ICRP(1991)	ICRP(2007)
Photons, all energies	1	1
Electrons and muons, all energies	1 \( \sum \times \)	1 J ×
Neutrons, energy < 10 keV	5	
10 keV to 100 keV	10	Continuous fnc
>100 keV to 2 MeV	20	of energy,
>2 MeV to 20 MeV	10	min 2.5, max 21
>20 MeV	5 -	)
Protons, other than recoil protons, >2 MeV	5	2 (also pions)
α particles, fission fragments, heavy nuclei	20	20

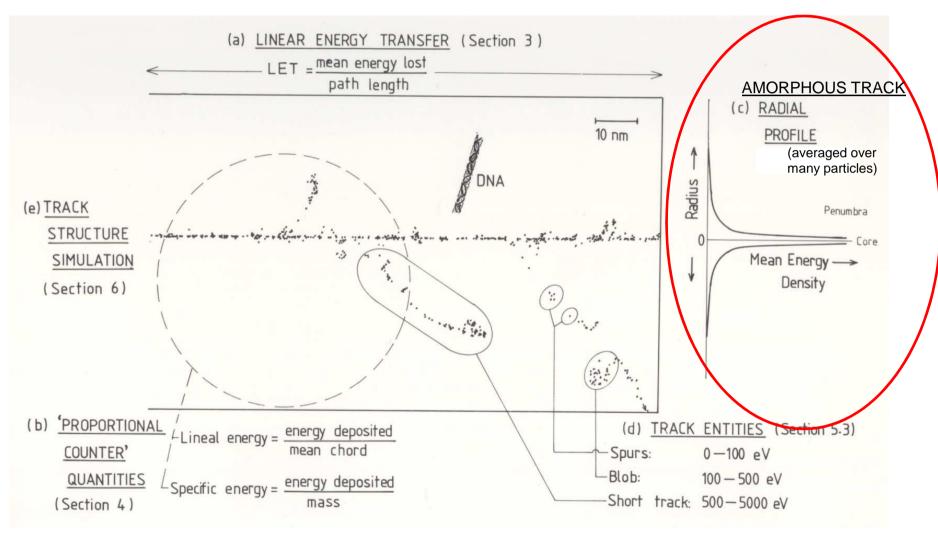
★ Implies equal risk per unit effective dose to body equivalent dose to a tissue > electron irradiations

absorbed dose to a tissue

For ALL photon and -- a major simplification

ICRP treats: absorbed dose from low-energy beta emitters (few keV) exactly as if from orthovoltage X-rays (~100 keV) or from high-energy gamma-rays (~ 1 MeV).

## Amorphous track: Radial dose distribution



## Amorphous track: Consider radial dose distribution

Indicates average lateral spread of particle track

( Averaged over many particles of same Z, E)

- BUT totally ignores stochastics of track
- Maximum width of track depends on particle velocity (V), i.e. on Energy/nucleon (not on Z)
- Ions of equal V, have  $\sim$  same <u>relative</u> track width and radial dose distribution ( $D_r \sim 1/r^2$ )
- Energy density in track depends on both Z and V

(as dominant term of Bethe-Block stopping power formula)

#### 1 MeV 1H Expt. ref. [39] 105 (27 keV/µm) Exc. 105 δ-rays 10 100 10 100 DNAt/ nm 1 µm 377 Mev/u <sup>20</sup>Ne (31 keV/µm) Expt. ref. [40] 103 Exc. 10 -ravs 10 100 104 109 105 10 DNA t/ nm 1 mm

Fig. 1a-b Comparison of calculations of radial dose distributions to experiments [39, 40]. Calculations show contributions from excitations and secondary electrons. a <sup>1</sup>H at 1 MeV (linear energy transfer, LET=27 keV/μm), b <sup>20</sup>Ne at 377 MeV/u (LET=31 keV/μm)

Adapted from: Cucinotta et al , Radiat Environ Biophys 38, 81-92 (1999)

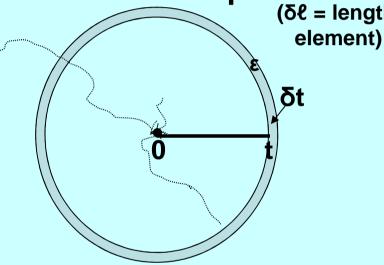
#### **Radial dose distribution**

Ave. Dose at radius t from ion path:

$$D(t) = \frac{\text{Energy}}{\text{Mass}} \text{ at radius t}$$

$$= \frac{\epsilon(t, \delta t)}{2\pi t \cdot \delta t \cdot \delta \ell \cdot \rho} \text{ deposited in element } \delta t \cdot \delta \ell$$

$$(\delta \ell = \text{length of } \delta t \cdot \delta \ell)$$



 $D(t) \sim 1/t^2$  over most of the profile

Note then that

 $\varepsilon(t) \sim 1/t$ 

i.e. <u>energy</u> falls off much more slowly than does <u>dose</u>

For heavy charged particles (i.e. protons and heavier):

With increasing distance r from track centre the local <u>dose</u>, D<sub>r</sub> falls off as

$$D_r \sim 1/r^2$$

Note: 1. Often misinterpreted as the track being too narrow:

Energy,  $\varepsilon_r$ , deposited in annulus at distance r, falls off much more more slowly, as

 $\varepsilon_r \sim 1/r$ 

Note: Mass of annulus increases proportionally with r

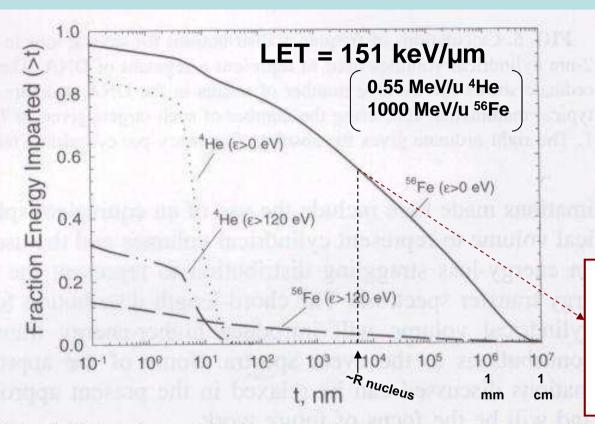
Hence,  $D_r \sim \epsilon_r/m_r \sim 1/r \cdot 1/r \sim \epsilon_r/r2$ 

2. Beware early descriptions based on "core and penumbra".

Misleading because:

~ 50 % of deposited energy was arbitrarily assigned to the "core" --- **not valid**; Definitions of "core" questionable.

#### Compare energy spread by ions of same LET



Fraction of energy imparted (>t):

$$\int_{t=t}^{t=\infty} \varepsilon(t).dt / \int_{t=0}^{t=\infty} \varepsilon(t).dt$$

For the high velocity
Fe ions:
nearly 60% of the track
energy escapes a
traversed cell nucleus

- FIG. 9. Calculations of the fraction of energy imparted in a nucleosome from radial distances greater than t. This fraction is shown for the total energy imparted (0 eV) and for the fraction of energy imparted above 120 eV from ions of identical LET: 151 keV/μm.
- i.e. Linear energy deposited within the cell nucleus is only
  - ~ 40% x 151
  - ≈ 60 keV/µm

Adapted from: Cucinotta et al, Radiat Res 153, 459-468 (2000)

#### Applications of amorphous track/radial profile

#### **Basic studies:**

Katz amorphous track structure model for effects of radiation on cells

Phenomenological model for radiobiological responses of cells to heavy charged particles.

Integrates radial dose profile for heavy ions with radiation response from gamma- or X-rays.

e.g. Katz et al, Radiat Res 47, 102-125 (1971)

#### **Radiation protection:**

• NASA risk model uses  $Z^{*2}/\beta^2$  instead of LET to specify Quality Factor, and paramaterizes NASA Quality Factor as a function of 'Katz' parameters ( $\Sigma_0$ ,  $\kappa$  and m).

(Cucinotta et al (2013) PLoS ONE 8(10): e74988.doi:10.1371)

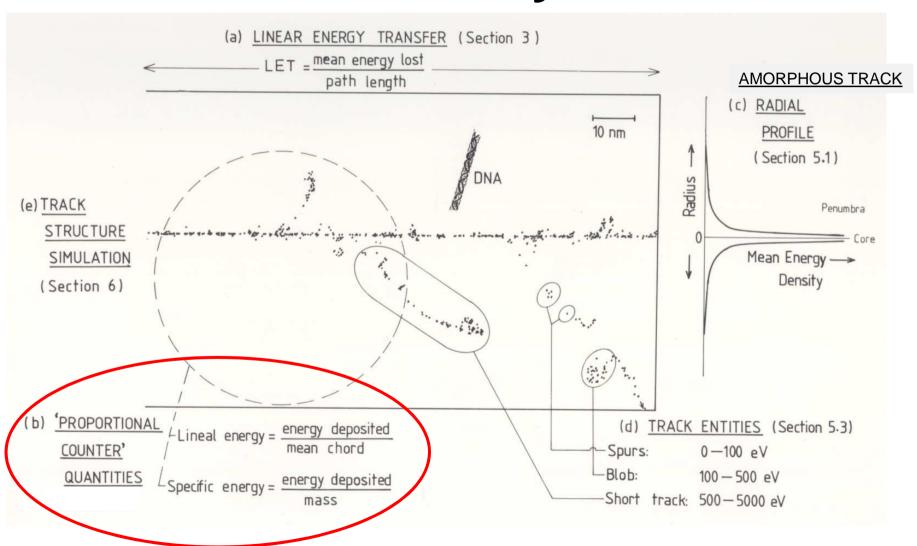
#### Radiotherapy:

Used in Local Effects Model (LEM) for radiotherapy with C ions (GSI)

Phenomenological model for cell killing by heavy ions. Conceptual similarities to Katz model.

e.g. Elsässer and Scholz, Radiat Res 167, 319–329 (2007)

# 'Proportional counter' microdosimetry



#### The Rossi Counter:

Low pressure tissue equivalent proportional counter (TEPC).

A major development of the 1950s & 60s



$$y = \frac{\varepsilon}{\overline{\ell}}$$
 (keV/µm)

Electric
Pulse ~ # ionizations
~ ε (energy imparted)

1/4" X 1" HELIX

ANODE

**Specific Energy:** 

$$z_1 = \frac{\varepsilon}{m}$$
 (Gy)

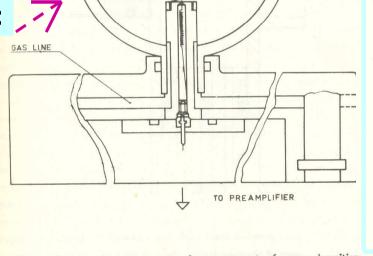


Fig. 1 The modified Rossi counter for measurements of energy deposition in small tissue-equivalent volumes.

From measured lineal energy spectrum of single tracks from neutrons (d,Be)

Measures <u>actual</u> stochastic events in a microscopic simulated tissue volume

### **Experimental Simulation of microscopic volume of tissue:**

Fill proportional counter with 'tissue-equivalent' gas at low pressure, such that that:

Energy loss for charged particle through counter gas
""" " " tissue volume

For scaling factor K: 
$$\frac{\rho_{gas}}{\rho_{tissue}} = \left[\frac{S_t}{S_g}\right] \left[\frac{p_g}{p_t}\right] = \frac{1}{K} \qquad \begin{array}{l} \rho = \text{density} \\ S = \text{stopping power} \\ p = \text{path length} \\ S_t/S_g \text{ approx} = 1 \end{array}$$

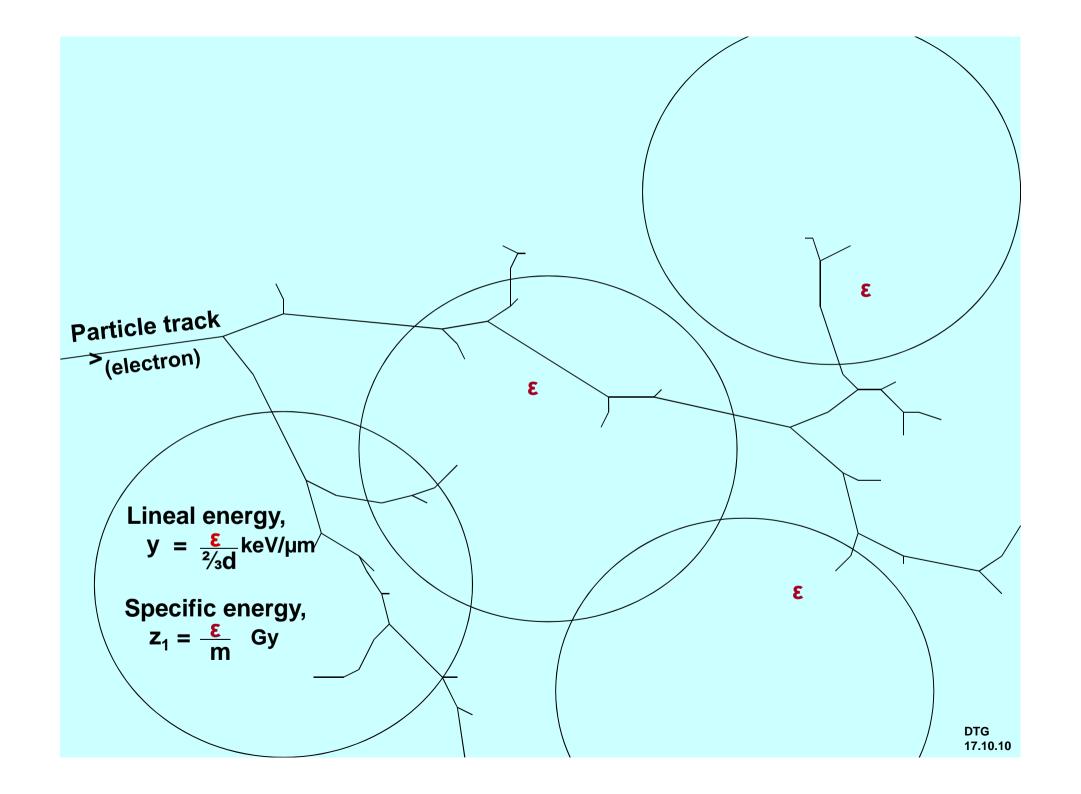
Typical scaling factor of 20,000 simulates 1 µm sphere in tissue with low-pressure gas in a 2 cm spherical prop counter

Most common is ~ 1  $\mu$ m simulation Also 0.5  $\mu$ m to 10  $\mu$ m (sub-nucleus to nucleus or cell sizes)

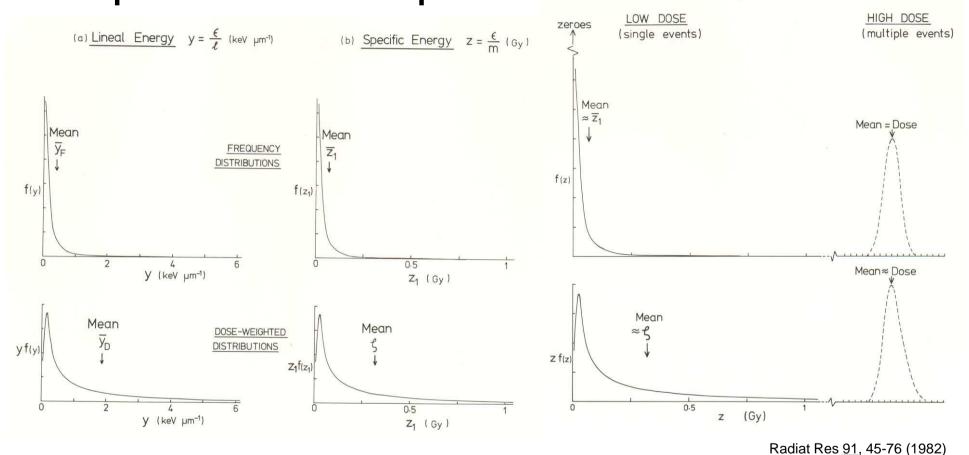
Practical limit of simulation: Down to tissue sites of ~ 0.3 µm diameter (still very large compared to DNA, nucleosomes, etc)

Later: Also solid state microdosimeters.

Much smaller volumes ('nanodosimetry').



Spectrum of lineal energy and specific energy for <sup>60</sup>Co γ-rays in sphere of diameter 1 μm



Obtained from experimental measurements with Rossi proportional counter.

Note: More usually plotted with log-scale abscissae (and ordinates therefore multiplied by y (or z) to preserve area normalization).

#### **Useful relationship**

### **Event frequency:**

$$\Phi = \frac{1}{\overline{z}_F}$$

gives average number of events ('hits') in the target volume per unit absorbed dose

Example: For a sphere of diameter 8  $\mu$ m in tissue irradiated with Co gamma-rays,  $\bar{z}_F = 1$  mGy (from measurements with Rossi counter).

Hence: For natural background radiation of 1 mGy per year, each cell nucleus of diameter ~ 8 µm is hit by radiation on average once per year

Approximations for irradiation with low-velocity charged particles of LET = L, (i.e. narrow tracks) crossing spherical targets:

$$y_F \approx L$$
  $z_1 \approx 0.204 \frac{L}{d^2}$   
 $y_D \approx \frac{9}{8} L$ 

$$\Phi \approx \frac{d^2}{0.204 L}$$

y, L in keV/µm z in Gy D in µm

#### Some Applications of y, z

• Hit-frequency evaluations:  $\phi = \frac{1}{\bar{z}_F}$ 

#### Basic radiobiology and risk modelling:

• Theory of Dual Radiation Action Developed hand-in hand w Rossi Counter during 1970s for radiobiology and cancer risk: Effect =  $\alpha D + \beta D^2 = k(D + z_D^2)$ Curr Top Radiat Res Q

8,85-158 (1972);
Radiat Res 75, 471-488 (1978)

Proposed as fundamental and mechanistic, but assumptions invalidated by experimental tests.

See: Radiat Prot Dosim <u>122</u>, 3-15 (2006)

Remains usable as phenomenological model for limited purposes.

Microdosimetric Kinetic Model (MKM) of cell death. [Hawkins, Int J Radiat Biol <u>69</u>, 739 (1996);
 Incorporates aspects of TDRA and other models as practical mathematical formalism.
 Radiat Res <u>160</u>, 61 (2003);
 Radiat Res <u>172</u>, 761 (2009) ]

#### **Radiation Protection:**

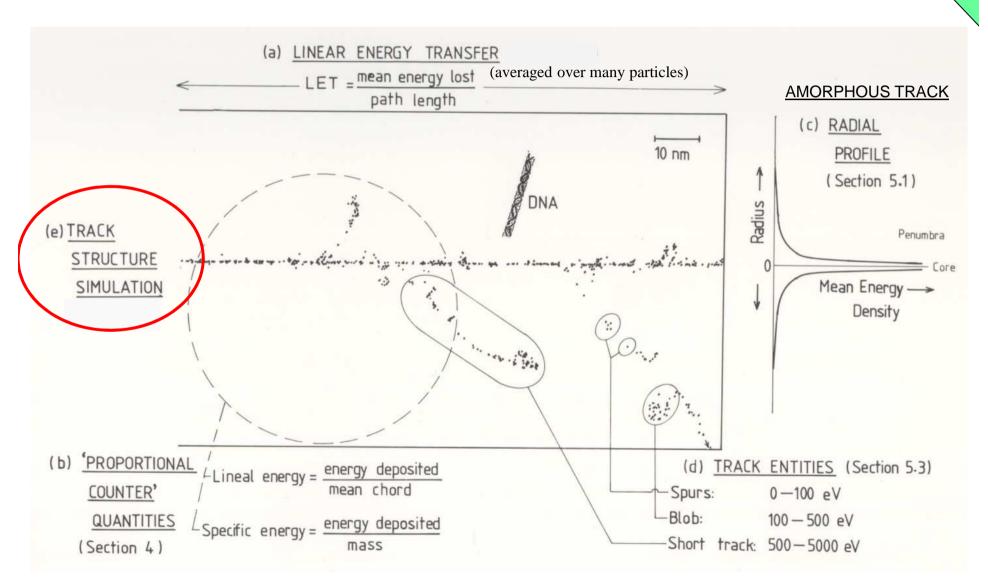
- (Task Group proposed specifying Q as fnc of mean lineal energy (y) instead of fnc LET, but never adopted by ICRP). (ICRU Report 14, 1986)
- <u>Wide</u> practical application in dosimeters: e.g. Measure y spectrum in mixed radiation field (prop counter and other devices), unfold as LET spectrum and hence evaluate Q(L) and equivalent dose rate.

#### Radiotherapy:

 Mathematical approach based on MKM for "biological dose" for planning of C-ion radiotherapy at HIMAC (Japan).
 Kase et al, Radiat Res 166, 629-638 (2006);

Sato et al, Int J Radiat Biol **88**, 143-150 (2012) Sato et al, Radiat Res **178**, 341-356 (2012)

## **Track structure:**



## Track structure

- Event-by-event simulations
- Provide ~ complete microscopic description of radiation
- BUT what to do with all the information ???

Reduce to the well-known microdosimetric/radiation-quality quantities?

Calculate 'novel' microdosimetric/radiation-quality quantities ??

Quantities on the nanometre scale (cf DNA, etc)?

#### Use for modelling:

- to provide new insights and generate new hypotheses on radiation mechanisms and effects
- to provide quantitative descriptions of known phenomena/data

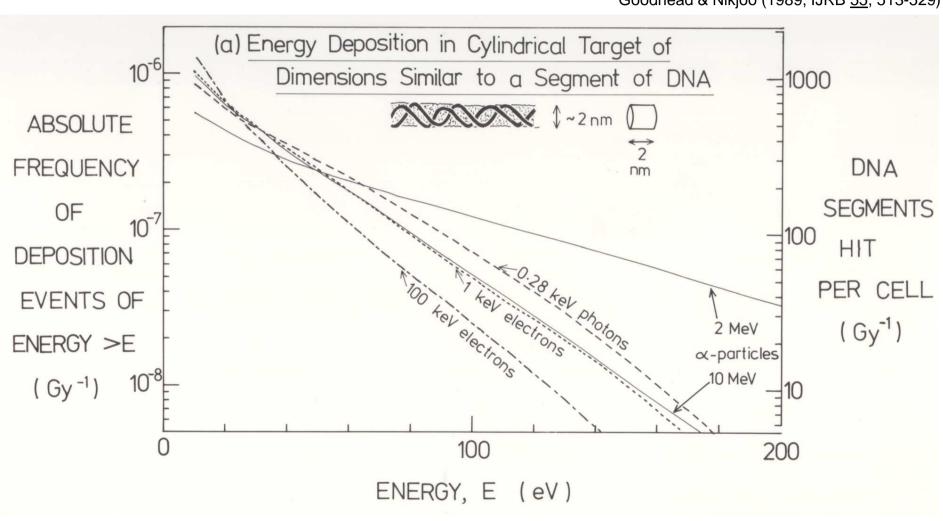
### **Modelling from track simulations**

1. A personal example:

**DNA Clustered damage** 

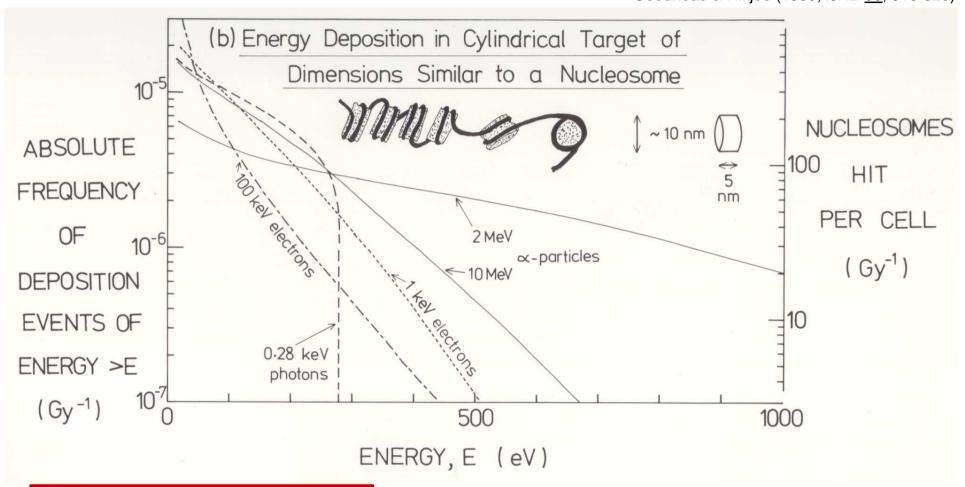
## Spectra of 'hit sizes' in DNA-sized targets from different radiation qualities (Calculated by sampling Monte-Carlo track-structure simulations)

Goodhead & Nikjoo (1989, IJRB 55, 513-529)



## Spectra of 'hit sizes' in nucleosome-sized targets from different radiation qualities (Calculated by sampling Monte-Carlo track-structure simulations)

Goodhead & Nikjoo (1989, IJRB <u>55</u>, 513-529)



Hypothesis of critical properties:

For Low-LET: ~ 100 eV in ~ 3-4 nm High-LET: ~ 300 eV in ~ 10 nm

## Frequency distribution of energy deposition, ε, in target volumes of interest for HZE\* exposures:

from deterministic model, which combines:
results from Monte-Carlo scoring of electrons
with average-track model of ions (amorphous track)

Cucinotta et al (2000) Radiat Res 153, 459-468

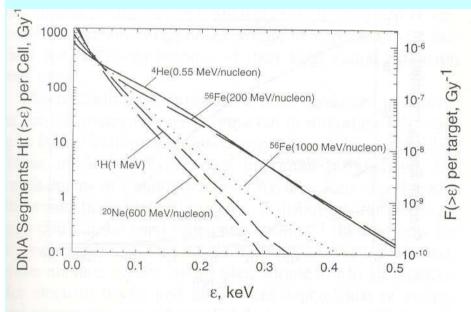


FIG. 5. Calculations of frequency distributions for several ions in  $2 \times 2$ -nm cylindrical volumes used to represent a segment of DNA. The left ordinate shows the average number of events in the DNA structure in a typical mammalian cell, using the number of such targets given in Table 1. The right ordinate gives the absolute frequency per cylindrical target.

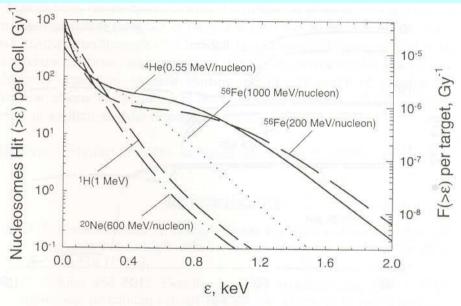
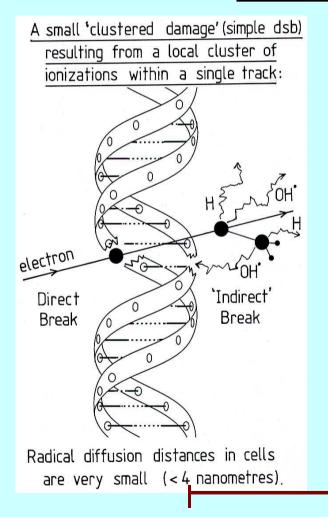


FIG. 6. Calculations of frequency distributions for several ions in a 10 × 5-nm cylindrical volume (nucleosome). The left ordinate shows the average number of events in the nucleosome structure in a typical mammalian cell, using the number of such targets given in Table 1. The right ordinate gives the absolute frequency per nucleosome cylindrical target.

- Well known that chromosome aberrations, and smaller mutations, can result from Double-Strand Breaks (DSB) in DNA
- Ionizing radiation is efficient at producing DSB
   because of <u>clustering</u> of ionizations within individual tracks



This simple Double-Strand Break has been produced by:

- one direct ionization, and
- one OH radical diffusing from an ionization in water very nearby ie Both were from a small

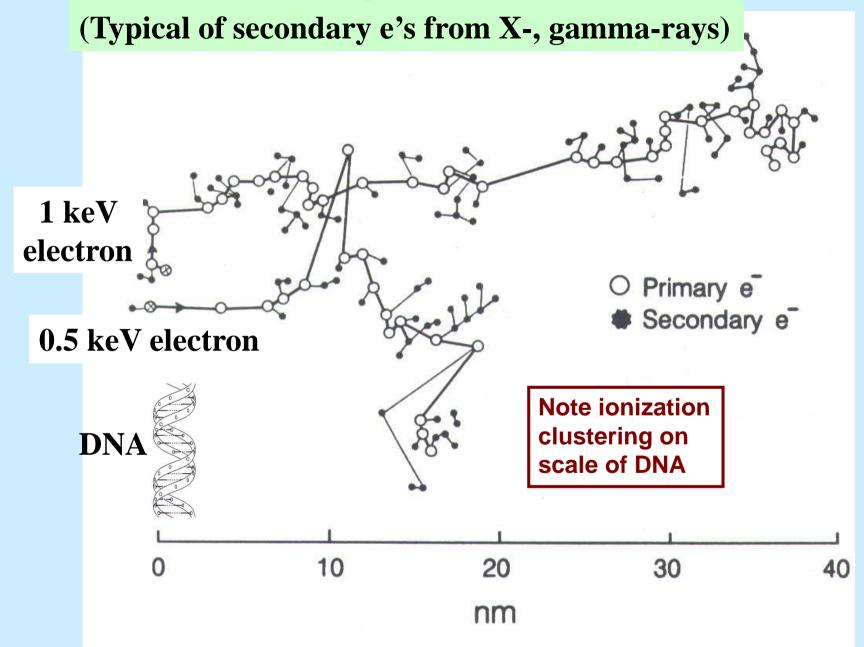
ie Both were from a small cluster of ionizations in a single electron track

#### Other DSB can be due:

- to two direct ionizations (ie Direct only)
- or to two OH radicals (ie Indirect only)

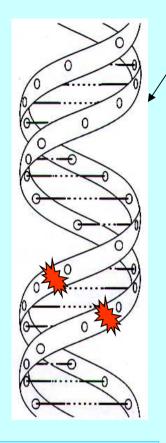
DSB result from <u>clustering</u> of ionizations on nm scale

## Two low-energy-electron tracks

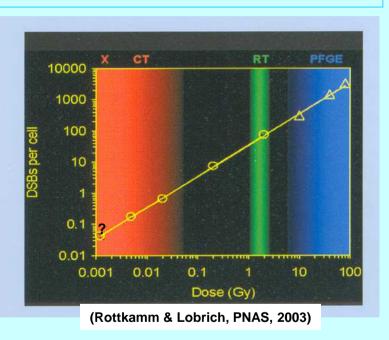


Example of Complex
Clustered Damage in DNA
resulting from a single
electron track from
low-LET radiation

2 nm



Damage in DNA resulting from a single electron track from low-LET radiation



**Complex DSB** 

Simple DSB

Yield of DSB is proportional to dose

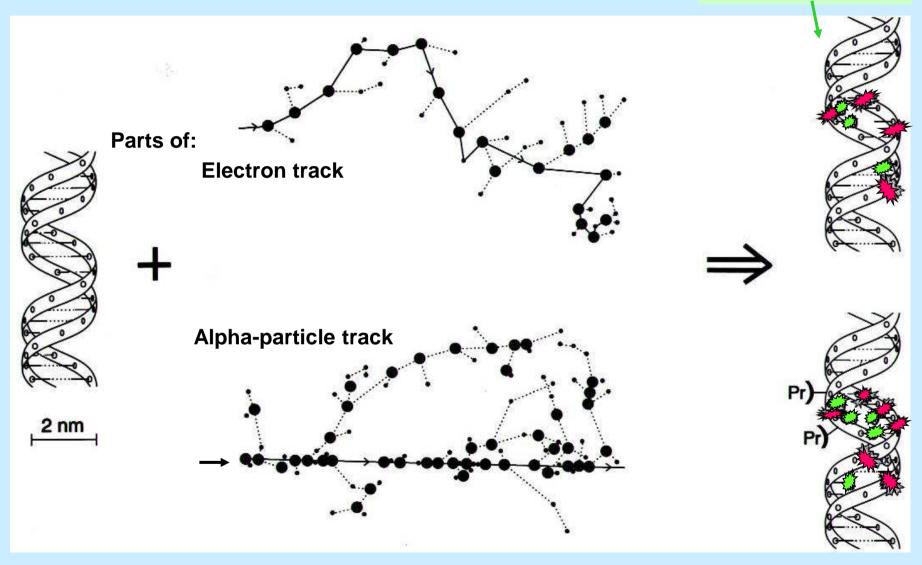
" number of tracks

Each DSB arises from a single track

**DSB = Double-Strand Break in DNA** 

## Single tracks of 'low'- LET or high- LET radiation can produce Complex Clustered Damage in DNA

Two examples of Complex Clustered Damage in DNA



### Clustered Damage in DNA

Simple damage (1 component):

Single strand break (SSB)



Damaged base (BD)



#### **Simple Clustered Damage (2 components):**

**Double strand break (DSB)** 



**Double base damage** 



SSB + BD



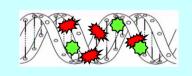
Also pairs on same strand

#### **Complex Clustered Damage** (3 or more components):

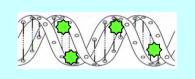
eg Complex DSB







Other combinations





Low-LET X, γ: ~ 20% of dsb are complex via 1 or more additional strand break(s)\*

~ 50% "

additional break(s) and/or base damage(s)\*

High-LET α:

" 1 or more additional strand break(s)\*

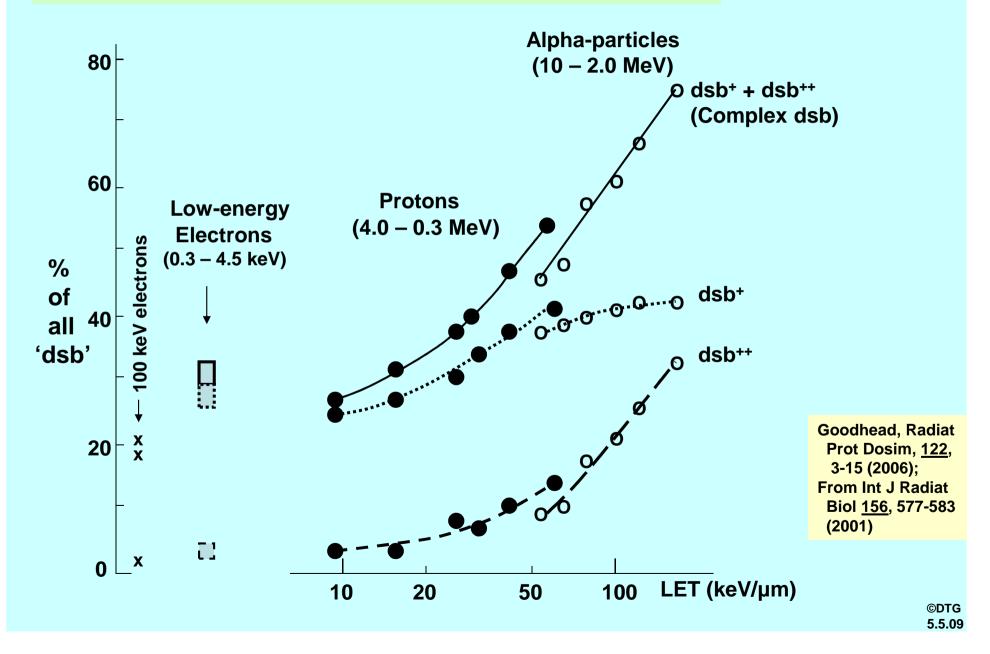
" additional break(s) and/or base damage(s)\*



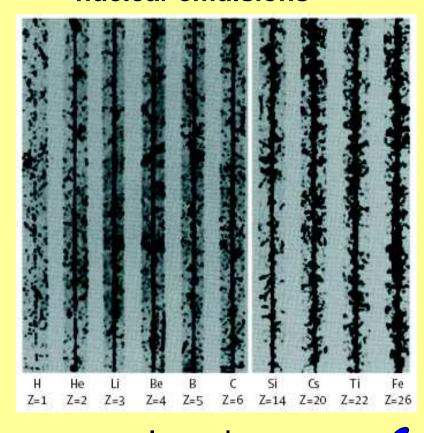
All radiations produce a substantial proportion of complex DSB

<sup>\*</sup> Nikjoo et al, Radiat Res <u>148</u>, 485 ('97); <u>156</u>, 577 ('02); IJRB 71, 467 ('97) 156, 577 ('02); Rad Prot Dosim 99, 77 ('02)

## The proportion of Complex DSB increases with LET. The degree of complexity increases with LET.



## Charged particle tracks in nuclear emulsions

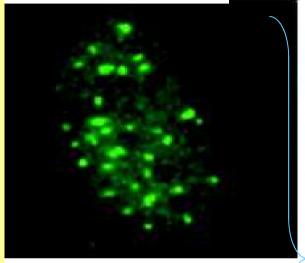


100 µm

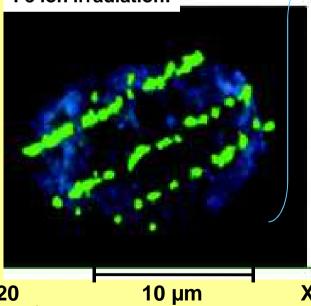
(Cucinotta & Durante, Lancet Oncol 2006)

## Fluorescent foci marking DSB in cell nuclei

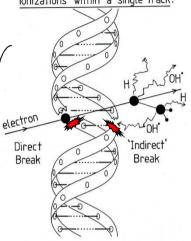
#### **Gamma-ray irradiation:**



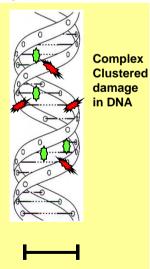
Fe ion irradiation:



A small 'clustered damage' (simple dsb)
resulting from a local cluster of
ionizations within a single track:



Radical diffusion distances in cells are very small (<4 nanometres).



2 nm

Magnification

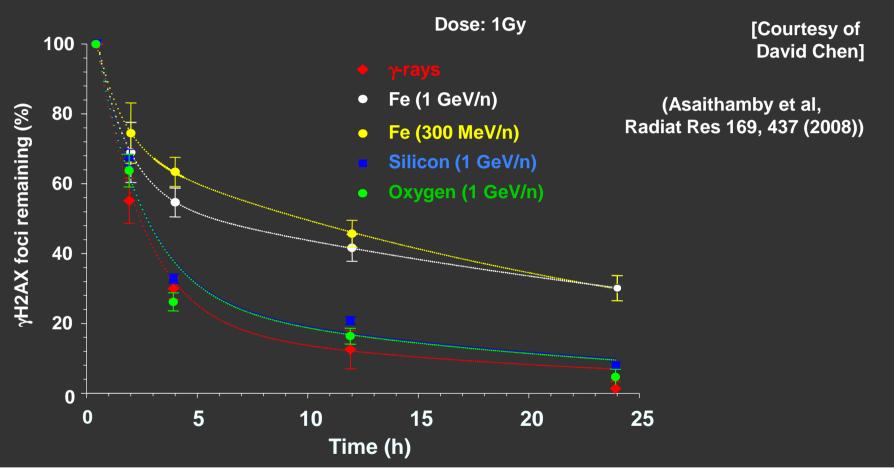
Cell nucleus

- - X 20

X 1600

DTG 4.2.14

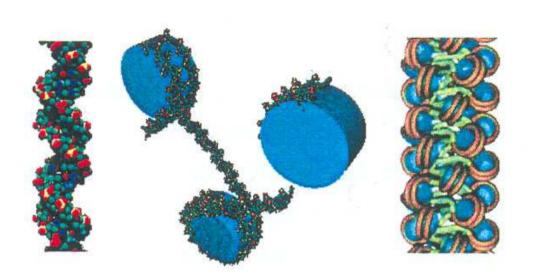
# Repair of DSBs induced by HZE particles in normal human skin fibroblasts



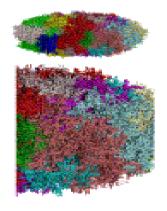
Such results are consistent with track structure predictions that there should be more-complex DSB from the more densely ionizing radiations (and hence more difficulty for repair)

### 2. Modelling at GSF/Helmholtz (Munich)

#### **Models of DNA organisation:**







Combine with M-C track-structure simulations to estimate damage from impact of tracks from radiations of many types



- DNA fragments,
  - Chromosome aberrations,
  - etc, etc

#### e.g. Friedland et al:

(2003) Radiat Res 159, 401 (2005) Radiat Phys Chem 72, 279 (2006) Radiat Prot Dosim 122,116 (2010) Radiat Res 173. 263 (2010) Radiat Res 173, 677 (2013) Mutat Res 756, 213

### **Closing comments:**

#### **Differences in radiation quality can lead to:**

- differences in <u>biological effectiveness</u> for the same quantity of radiation (e.g. the same absorbed dose) --- can quantify ~as RBEs
- qualitative differences in biological effects --- cannot use scaling to specify

#### **Effects of internal emitters depend on**

- Dose localization/inhomogeneity
   AND
- Radiation quality

#### Practical attempts to account for radiation quality include:

Radiation protection (very approx.):  $w_R$  (radiation weighting factor)

or Q (quality factor) as function of LET

More detailed risk assessments: Best available information on specific RBEs

NASA astronauts' risk model: QF as function of  $Z^{*2}/\beta^2$ 

Therapy, non-cancer effects, etc: e.g. Estimate 'Gy-Equivalent' doses for the

system

All have substantial short-comings ---- Much research to be done!!!



### **THE END**