



UK Health
Security
Agency

Biological and physical retrospective dosimetry

**16th EURADOS School: Contribution of dosimetry in the field of nuclear emergency preparedness and radiological accident management,
June 2023**

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Dose assessment approaches

PHYSICAL DOSIMETRY



Dosimetry badge

Fortuitous: Luminescence
Electron spin resonance
Bioassays

BIOLOGICAL DOSIMETRY



Cytogenetics
(Dicentric, FISH, PCC, MNA)#
Biochemical markers
Somatic mutations
Gene/miRNA expression

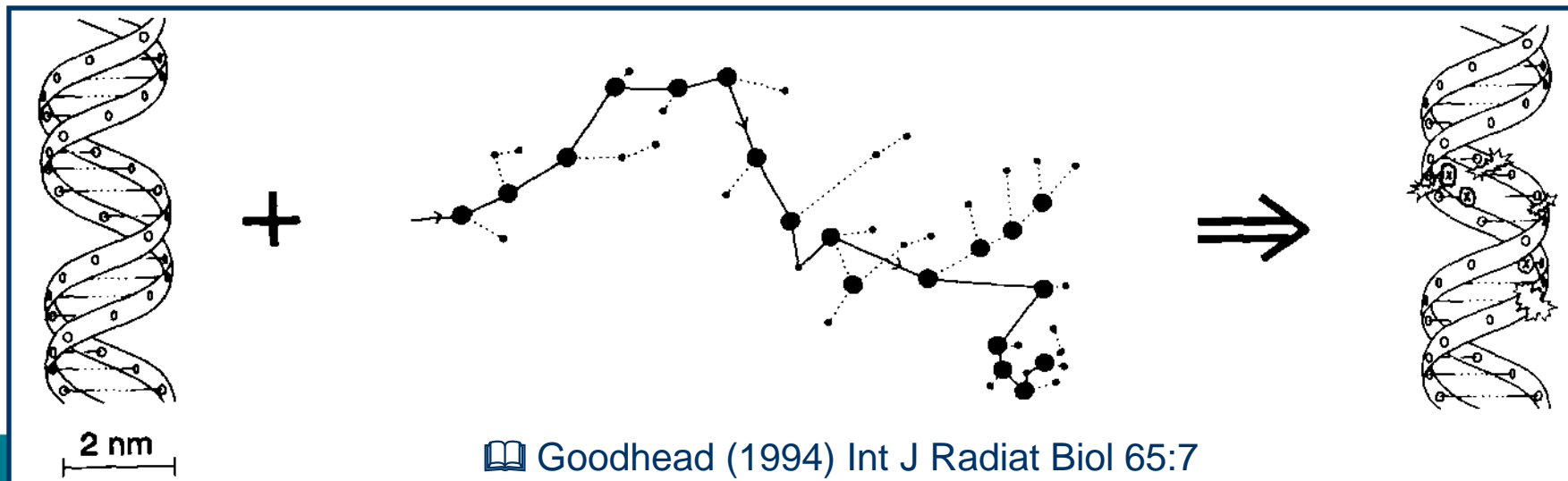
CLINICAL DOSIMETRY



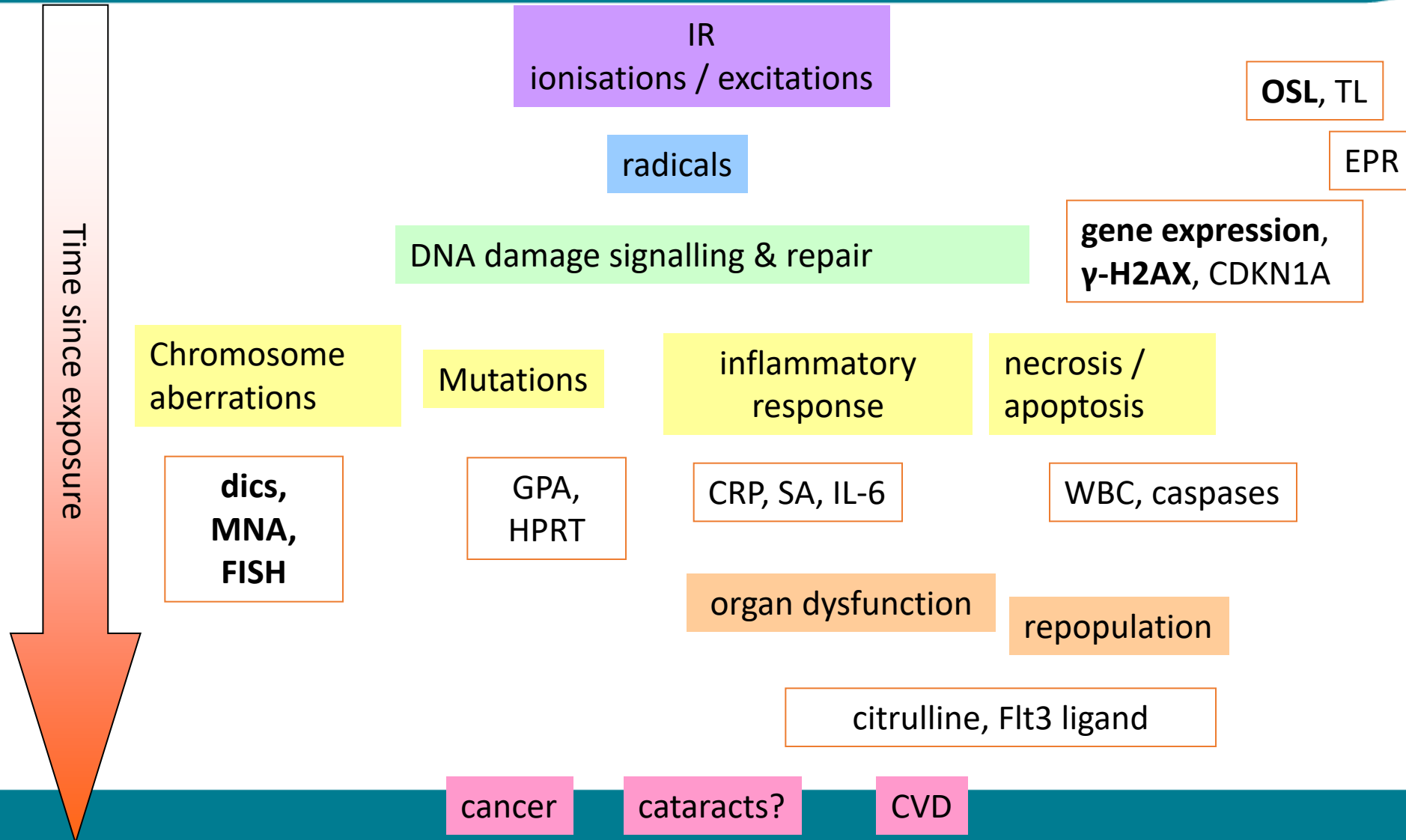
Vomiting
Diarrhoea
Blood cell count
Skin reactions...

Spectrum of damage induced by IR

| | | |
|--|-------------|--|
| Damage induced by 1 Gy X-rays in a human cell: | physical | <ul style="list-style-type: none">• 100,000 ionisations in the cell nucleus• 2,000 direct ionisations in the DNA |
| | biochemical | <ul style="list-style-type: none">• 1,000 single-strand breaks• 1,000 damaged bases• 150 DNA protein crosslinks• 35 double-strand breaks |
| | cellular | <ul style="list-style-type: none">• 0.1 dicentrics / micronuclei• 0.3 lethal events• 10^{-5} <i>hprt</i> mutations |



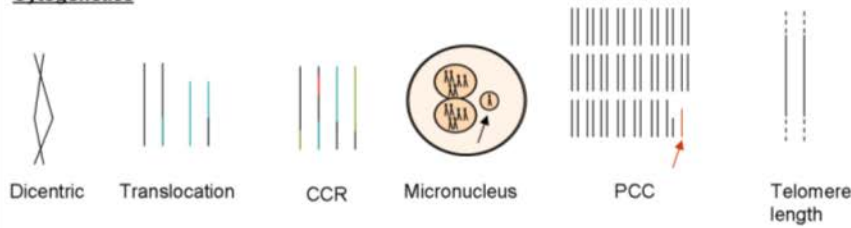
Biological effects of IR -> Biodosimetry



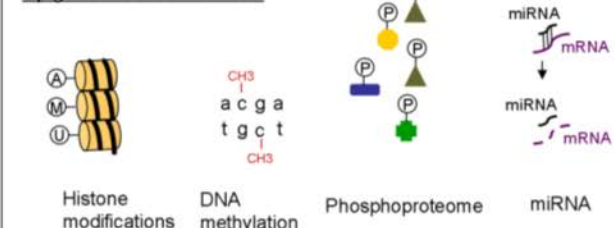
Exposure biomarkers

Pernot et al, Mutat Res 751: 258-286

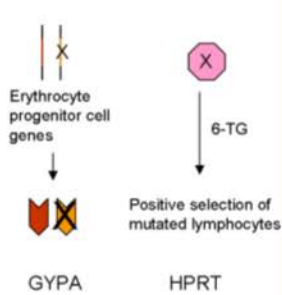
Cytogenetics



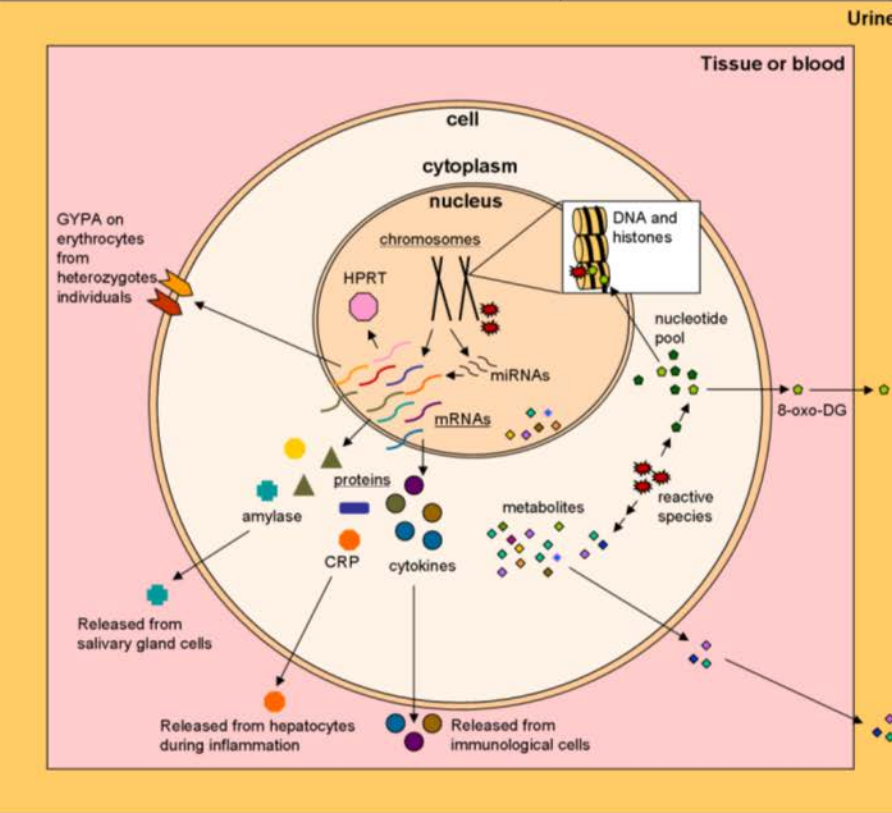
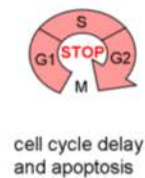
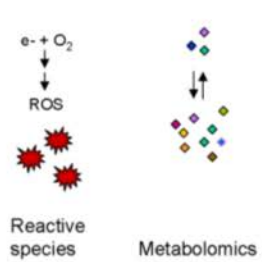
Epigenomic modifications



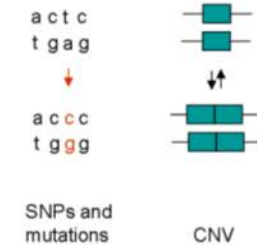
Induced mutations



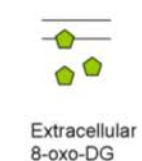
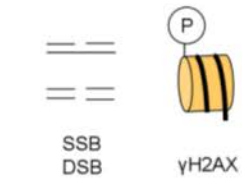
Others



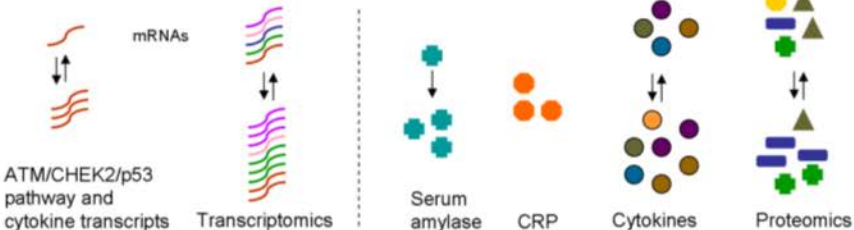
Germline inherited mutations or variants



DNA damage and nucleotide pool damage



Transcriptional and translational changes



The ideal dosimeter

Specific to radiation

Persistent effect

Low background

Ease of sampling

Low donor variability

Rapid analysis

Low doubling dose

Low cost

Dose response calibration

'Risk meter'

Characteristics of biodosimetry methods

📖 Ainsbury et al (2011) Radiat Prot Dosimetry, 147:573-92; Rothkamm & Lloyd (2013) Comprehensive Medical Biophysics Vol. 8, Ch.14; Wojcik et al, 2017, IJRB.

| Method | Radiation-responsive tissue | Time since exposure: days months years | Exposure: Protracted Non-uniform | Time (h) from sample receipt to dose estimate | Agent specificity | Dose range (Gy) for photon equivalent acute whole body exposure 24 h ago [°] | Triage use | Automated analysis | Standardisation |
|---------------------|--------------------------------|---|--|--|-------------------|--|------------|--------------------|------------------------|
| OSL/EPR | PED [‡] | d | P | <1 | IR | >0.01 | ✓ | Yes/No | ISO in preparation |
| OSL | Teeth / nails | d | | <1 | IR | >3 | ✓ | Yes | |
| EPR | Teeth / bones | d – m – y | P | 1-48 | IR | >0.05/>1 | | No | |
| γ-H2AX | Lymphocytes | d | N | 3 | Genotoxins | >0.2 | ✓ | Yes | ISO in preparation |
| Gene expression | Peripheral blood | d | N | 4 / 36 ^{&} | Genotoxins | >0.1 | ✓ | Yes | |
| Dicentrics – full | Lymphocytes | d – m | P; N | 55 | IR | 0.1 to 5 | | Semi | |
| Dicentrics - triage | Lymphocytes | d – m | P | 52 | IR | 0.5 to 5 | ✓ | Semi | ISO 19238 ISO 21243 |
| PCC fragments | Lymphocytes | d | N | 2 [§] | IR | 0.2 to 20 | ✓ | No | ISO in preparation |
| PCC rings | Lymphocytes | d – m | P | 40 [*] | IR | 1 to >20 | ✓ | No | |
| Micronuclei | Lymphocytes | d – m | P | 75 | Genotoxins | 0.2 to 4 | ✓ | Yes | |
| MN centromere | Lymphocytes | d – m | P | 80 | Genotoxins | 0.1 to 4 | | Semi | |
| FISH | Lymphocytes & their stem cells | d – m – y | P | 120 | IR | 0.25 to 4 | | No | |
| HPRT | Lymphocytes | d – m | | 400 | Mutagens | >1 | | No | |
| GPA | Erythroblasts | m – y | | 3 | Mutagens | >1 | | Yes | |
| TCR | Lymphocytes | d - m | P | 180 | Mutagens | >0.5 | | Yes | |
| CRP | Hepatocytes | d | | 1 | Wide range | >1 | ✓ | Yes | Routine diagnostics |
| SA | Salivary gland | d | | 1 | Wide range | >1 | ✓ | Yes | Routine diagnostics |
| Flt3 ligand | Bone marrow | d | | 1 | Wide range | >1 | ✓ | Yes | Routine diagnostics |
| Citrulline | Enterocytes | d | | 1 | Wide range | >1 | ✓ | Yes | Routine diagnostics |
| Metabolomics | Range of organs | d | | <1 | IR | >1 | ✓ | No | |

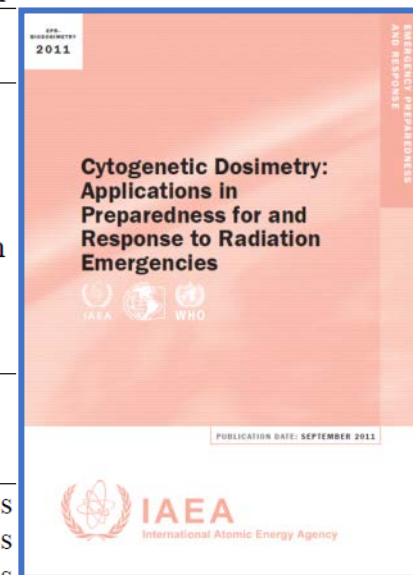
[‡]Personal electronic devices

[§]PCC fusion method

^{*}PCC chemically induced

[&]PCR / array analysis

[°]Longer post-exposure time required for GPA to allow for mutant erythroblasts to mature to erythrocytes



Principles of retrospective dosimetry

1. **Identify** potentially exposed individuals

2. Take a **sample**

A. Blood for biodosimetry

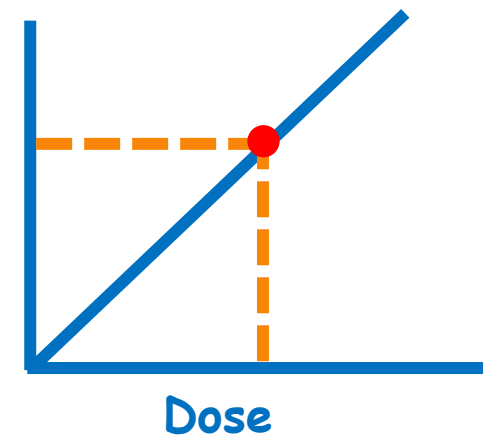
B. Personal belongings for OSL/EPR



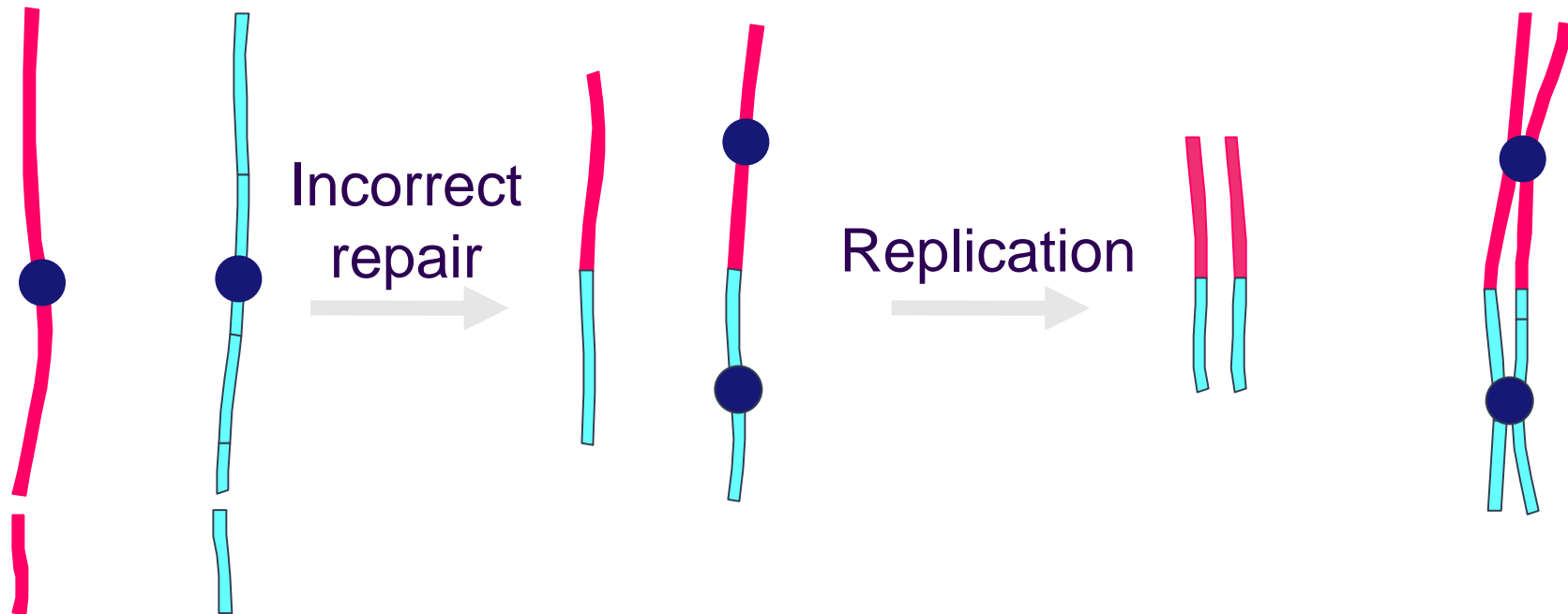
3. **Measure** radiation-induced signal

4. **Interpret:** Compare the test result with a **Yield** calibration curve for dose response; estimate uncertainties

5. Inform **clinical treatment** decisions

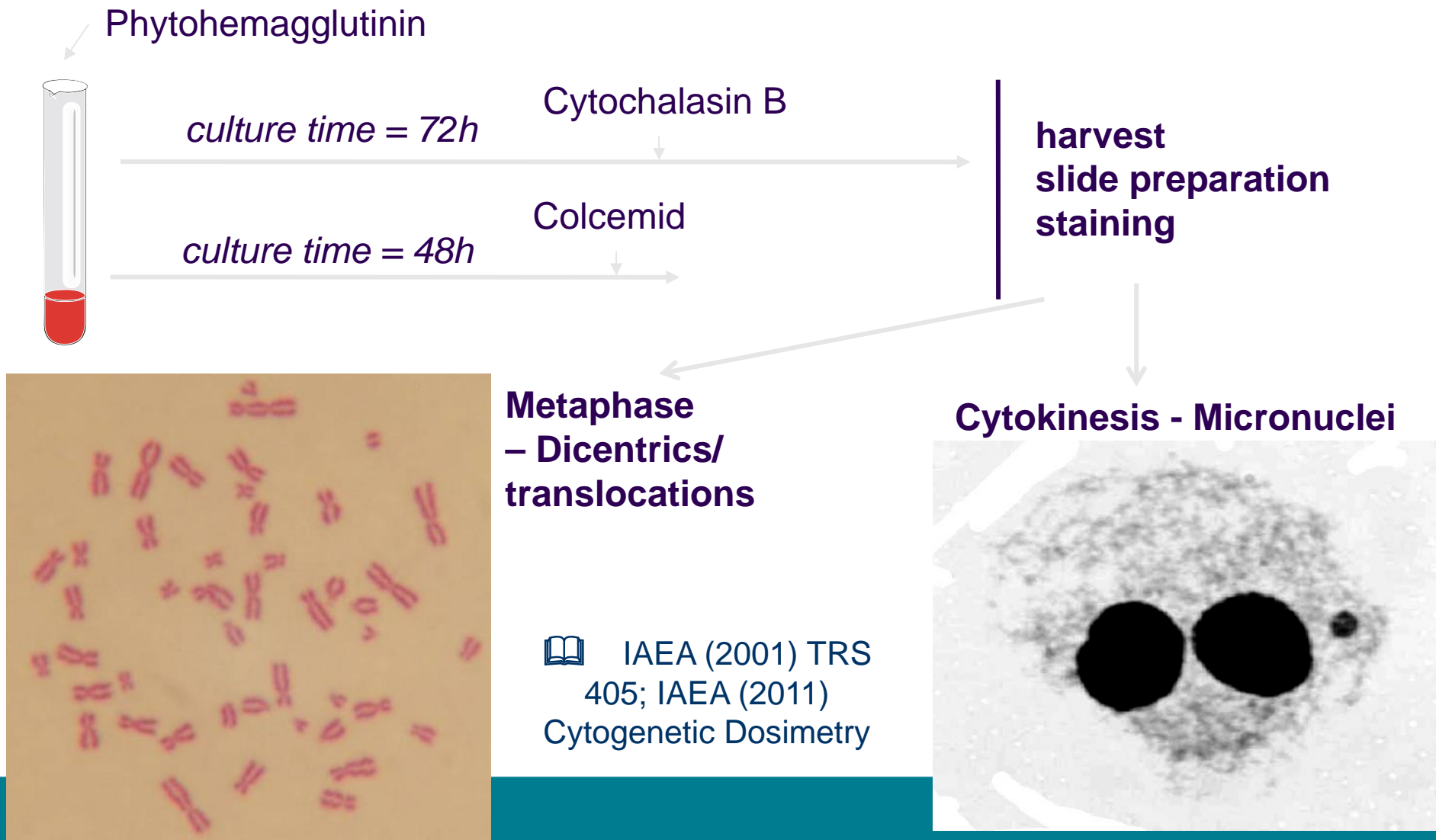


Dicentric formation



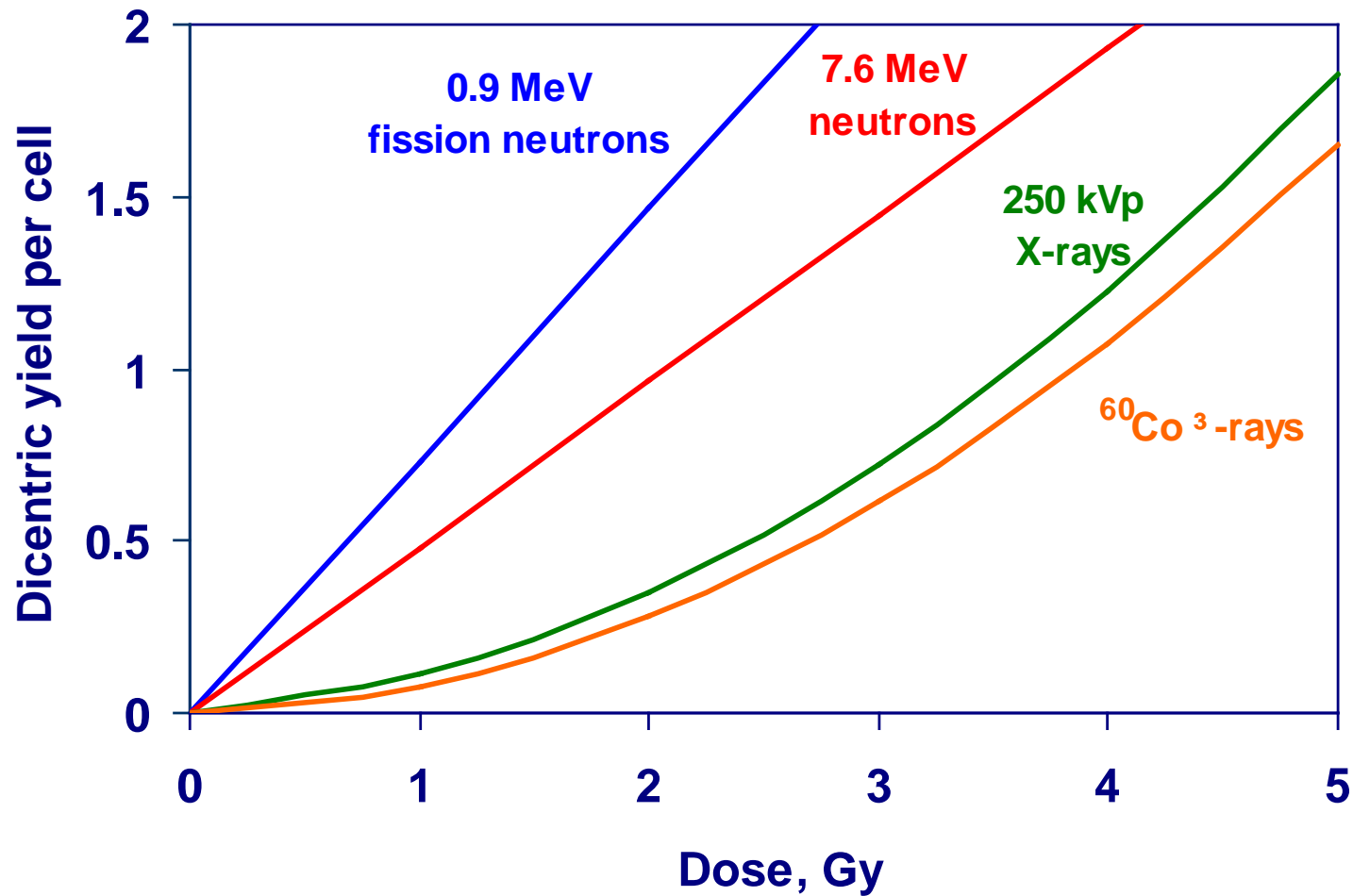
Chromosome dosimetry

- Blood lymphocyte culture



Dicentric - Dose response relationship

Human lymphocytes:



Automatic dicentric hunting

📖 Vaurijoux et al (2009) Radiat Res 171:541

Manual scoring:

- ~ 50 cells in 1 hr
- ~ 500 cells in 10 hr

Automatic scoring:

1. Rapid response mode:

- 150 cells in 30 mins; ~ 2 min staff effort;
- +/- 0.5 Gy

2. Full mode:

- 3000 cells in 10 h;
- ~ 40 min staff effort;
- +/- 0.15 Gy

The screenshot displays the Metafer4 software interface. The main window shows a chromosome spread with two red boxes highlighting dicentric chromosomes. The right-hand panel features the 'Metafer 4 AutoCapt' logo, version 'V3.1.3', and a date/time stamp '04.06.2007 16:55:32 swe'. Below this is a table with columns for 'CDics', 'NClas2', 'NObj', 'AuxVar6', 'ADics', 'NClas3', 'NChrs', 'AuxVar5', 'AuxVar1', 'AuxVar2', 'AuxVar3', and 'AuxVar4'. The bottom status panel shows 'AFD38E3g~A' with a count of 976, and a table with columns for 'Undefined', 'Marked', 'Rejected', 'Deleted', 'Cell No', 'Quality Rank', 'Quality Score', 'Cell ID', 'Status', and 'Class'.

| CDics | NClas2 | NObj | AuxVar6 |
|-------|--------|------|---------|
| 3 | 3 | 47 | 0 |

| ADics | NClas3 | NChrs | AuxVar5 |
|-------|--------|-------|---------|
| 3 | 3 | 25 | 0 |

| AuxVar1 | AuxVar2 | AuxVar3 | AuxVar4 |
|---------|---------|---------|---------|
| 0 | 0 | 0 | 0 |

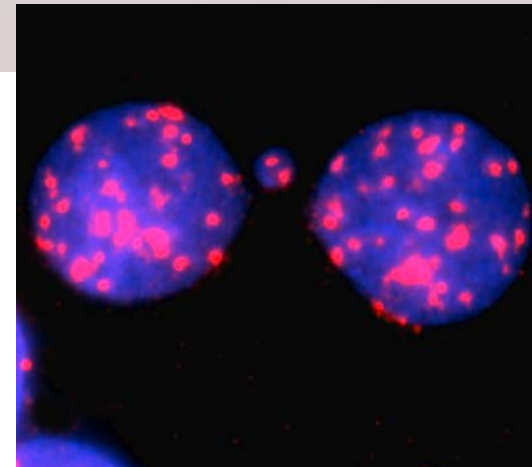
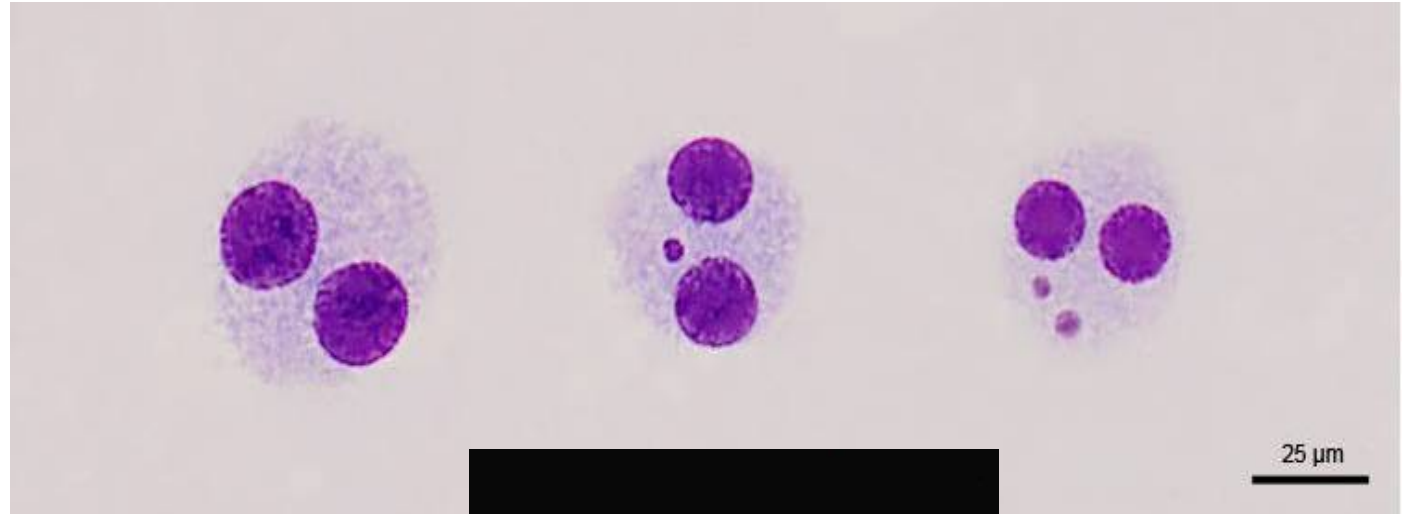
| Undefined | Marked | Rejected | Deleted | Cell No | Quality Rank | Quality Score | Cell ID | Status | Class |
|-----------|--------|----------|---------|---------|--------------|---------------|---------|--------|-------|
| 0 | 776 | 200 | 0 | 282 | 327 | 83 | 501 | Marked | O.K. |

The micronucleus assay

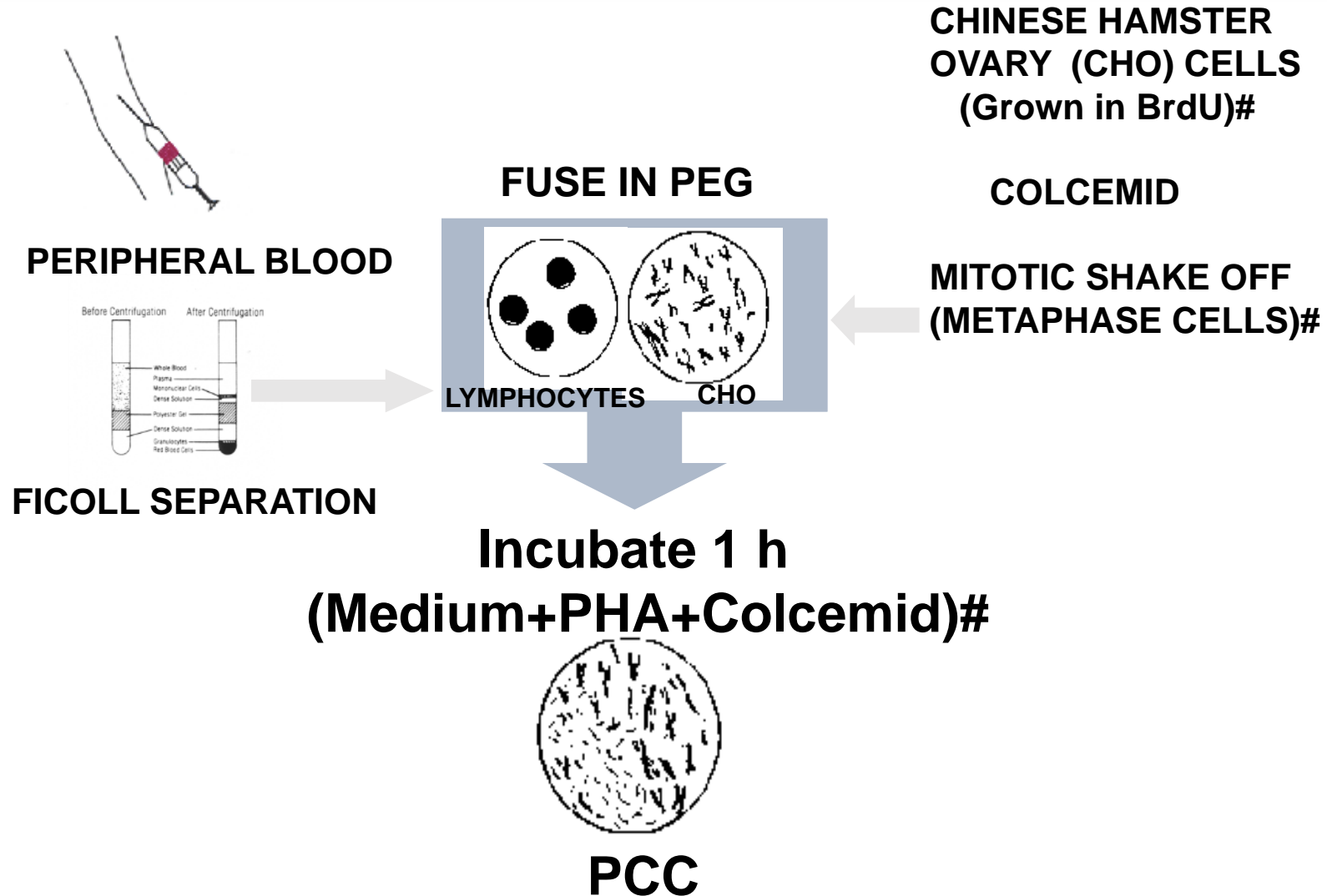
📖 Vral et al (2011) Mutagenesis 26:11

Comparison to dicentric assay

- MN arise mainly from acentric fragments
- Scoring easier & faster
- Not specific to radiation
- More background noise (especially for women...)
- Less sensitive (dose limit >0.2 Gy)
- Unstable signal fades like the dicentric
- Cannot easily discriminate between total and partial body exposure



Premature chromosome condensation



Some limitations with current cytogenetic analysis methods



Vinnikov et al (2010) Radiat Res 174:403

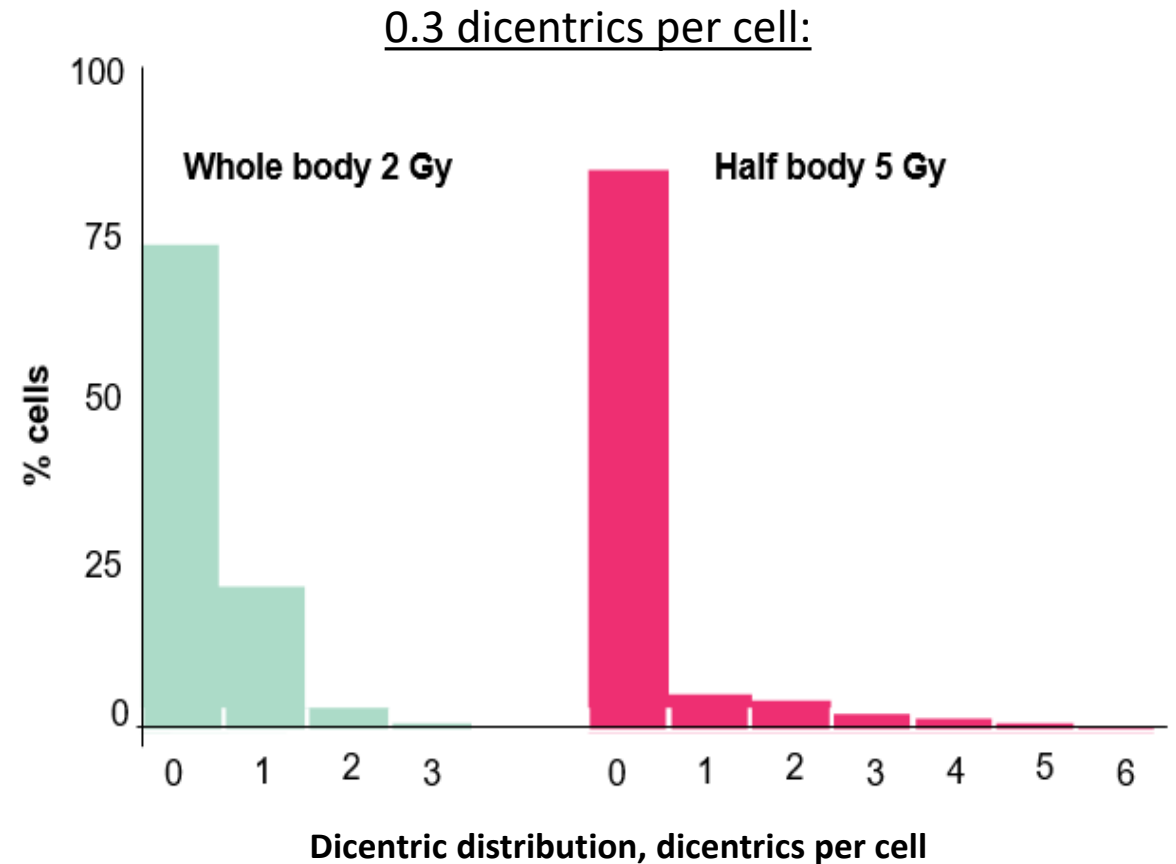
Ideal scenario: Acute, whole body, homogeneous, external irradiation of a known low LET source, with dose in the region of $> 0.1 - \sim 5$ Gy...

Real life exposures:

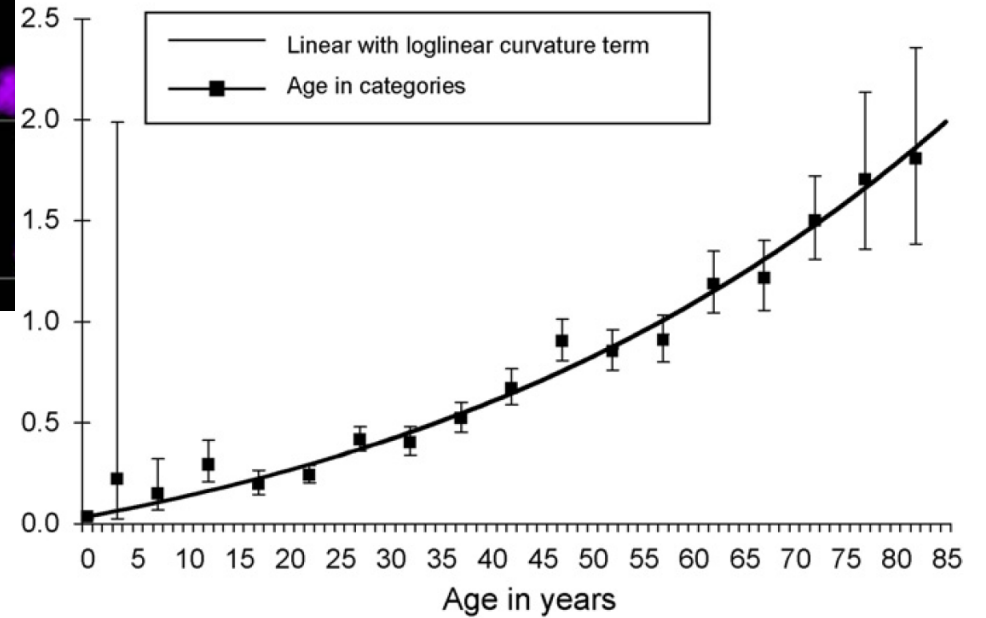
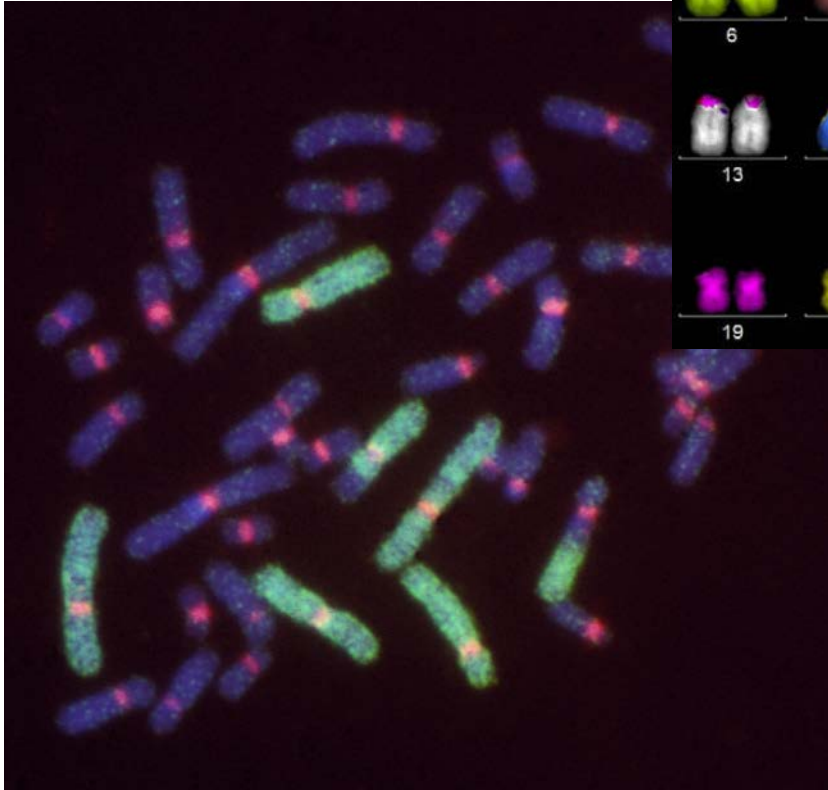
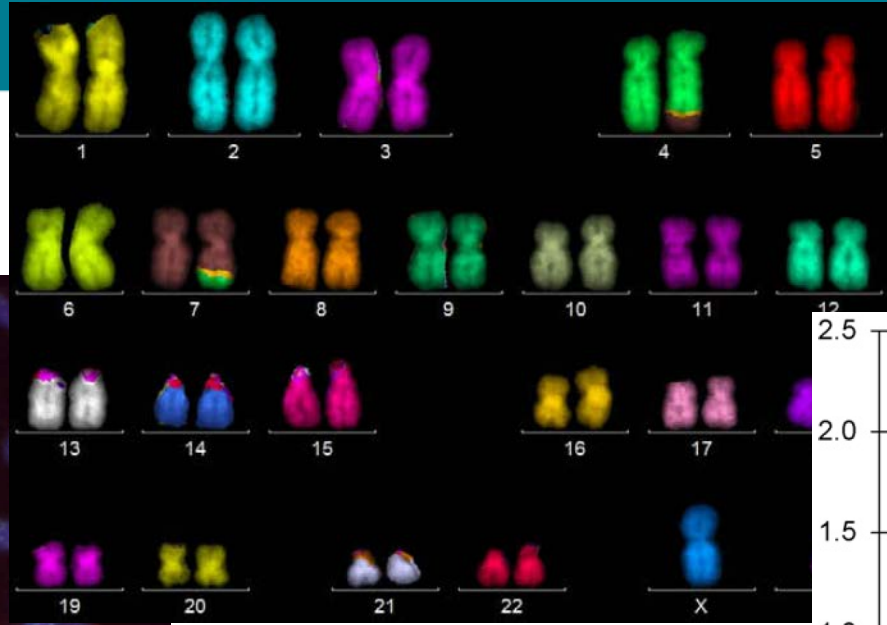
- Delayed blood sampling
- Protraction or fractionation, chronic exposures
- Inhomogeneity
- Very high or very low doses
- High LET radiation
- Internally deposited radionuclides
- Mass casualty scenarios
- Inter-scorer, -lab, -assay variation and other confounders

Non-uniform exposure

- Practically all accidents involve inhomogeneous / part-body exposure
- Dicentrics in lymphocytes indicate averaged whole body dose
- Uniform exposure (e.g. in test tube) – Poisson distribution
- Non-uniform – Overdispersion
- Dolphin et al.: ‘Contaminated Poisson’ method



Fluorescence *in situ* hybridisation for stable translocations



📖 Edwards et al (2005); Edwards et al (2007);
Hartel et al; Sigurdson et al (2008)

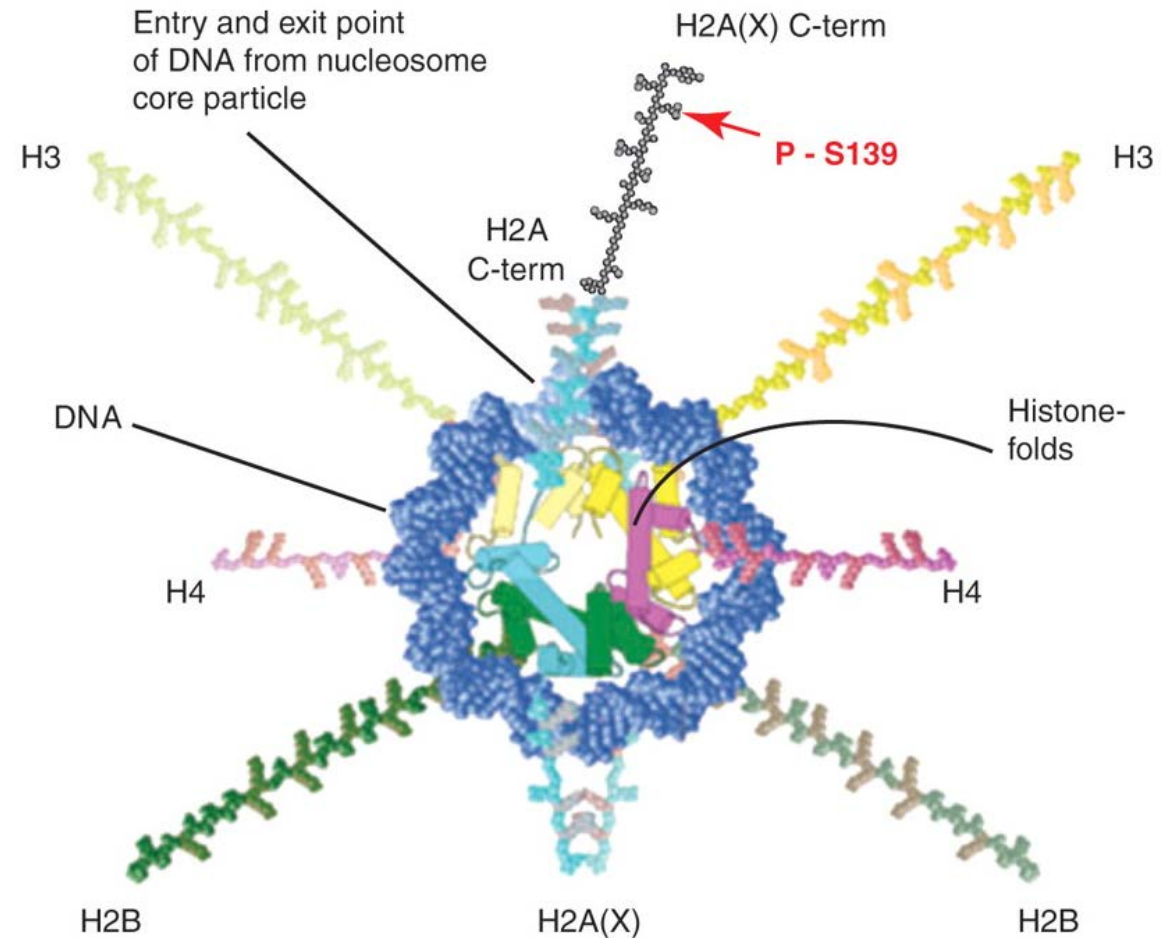
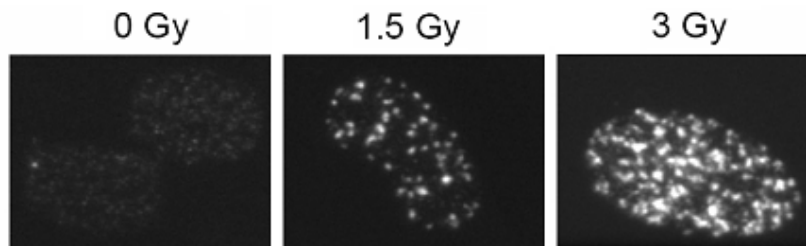
DNA Damage - Gamma-H2AX as a marker for DSB

📖 Rothkamm & Horn (2009), Ann Ist Super Sanita 45:265-71; Barnard et al (2013), Genome Integr 4:1

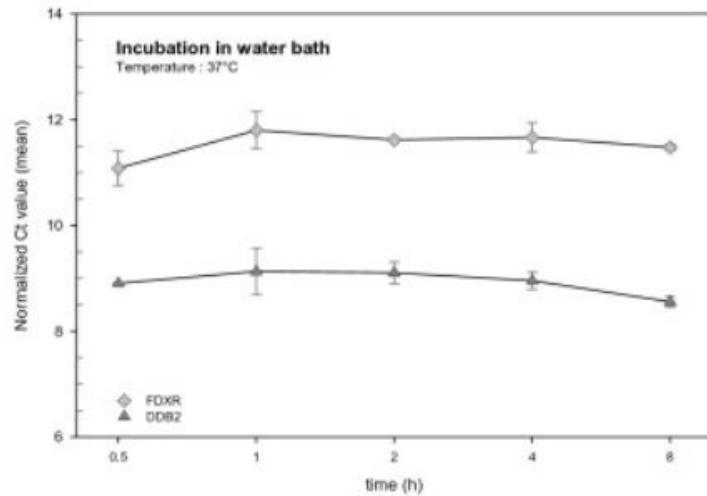
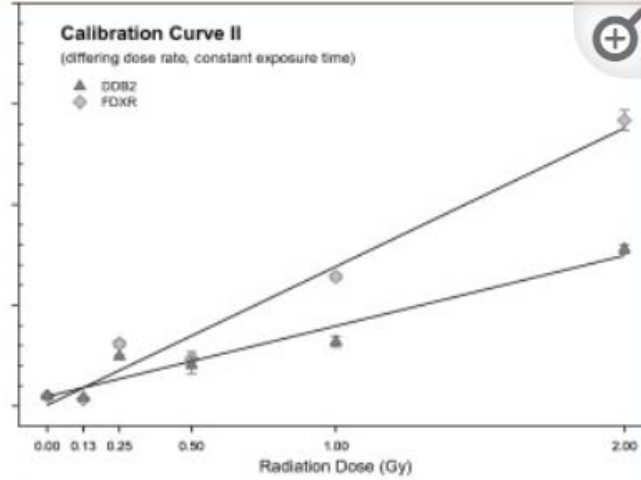
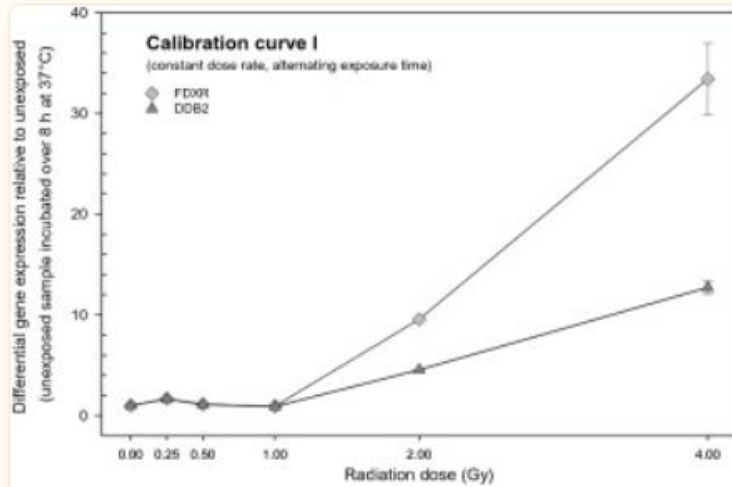
- H2AX is a component of the DNA packaging scaffold (chromatin) in the cell

- Each radiation-induced DSB induces hundreds of H2AX phosphorylations.

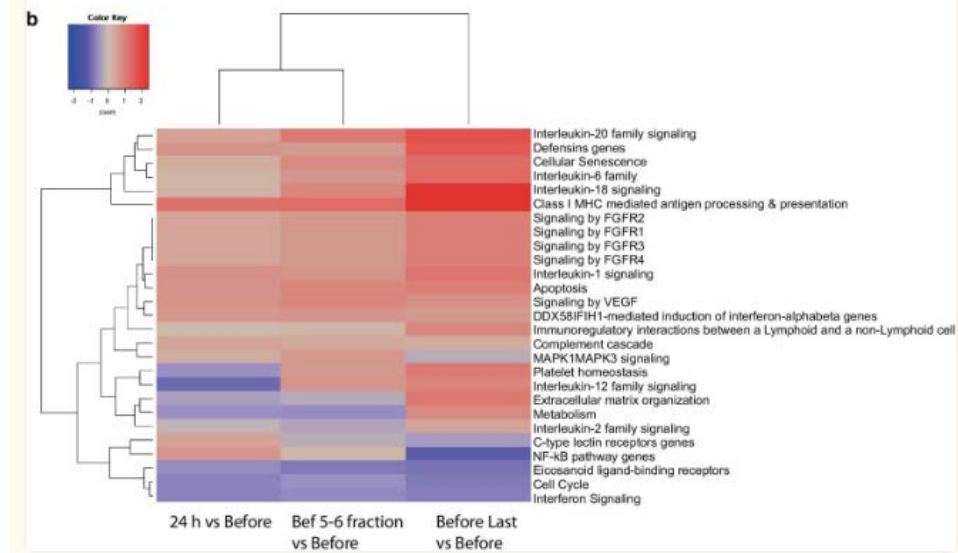
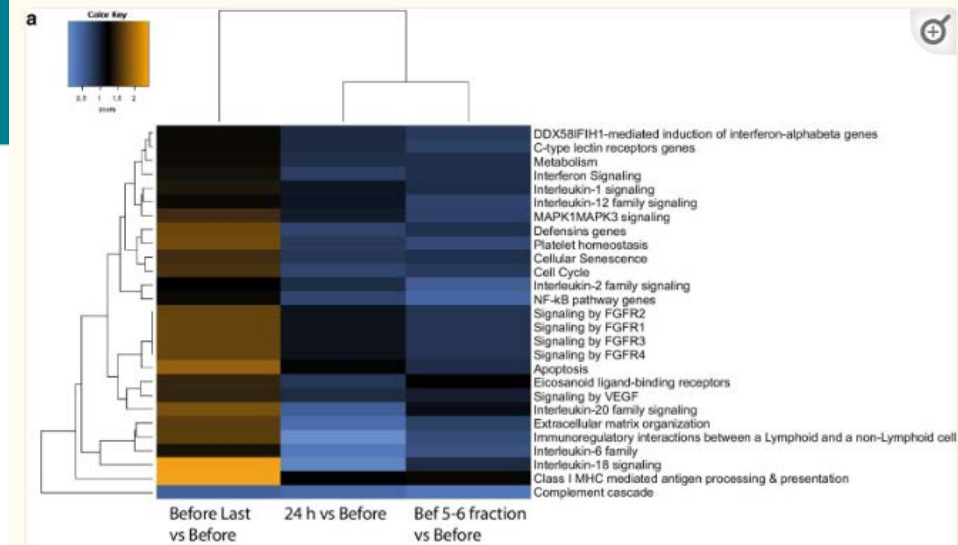
- γ -H2AX antibody visualises and quantifies individual DSBs at very low doses.



Gene expression, transcriptional responses

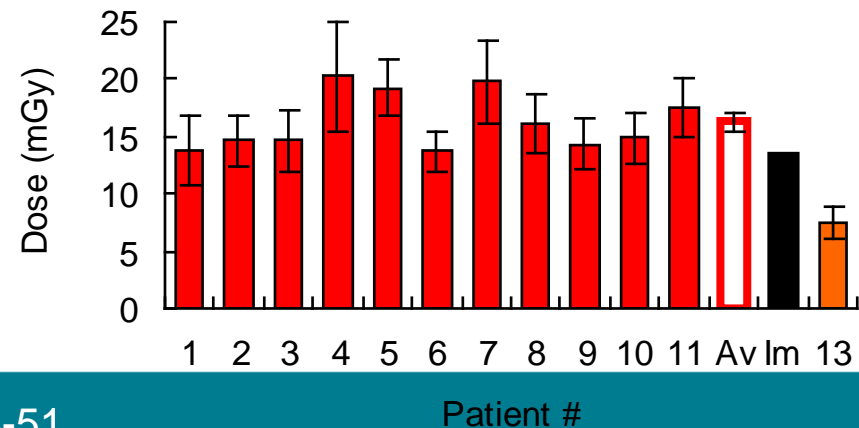
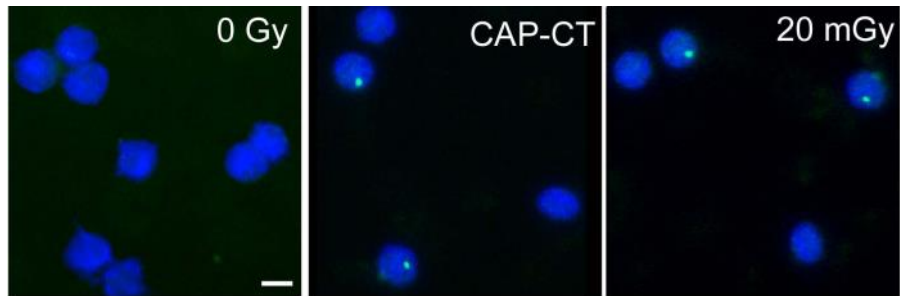
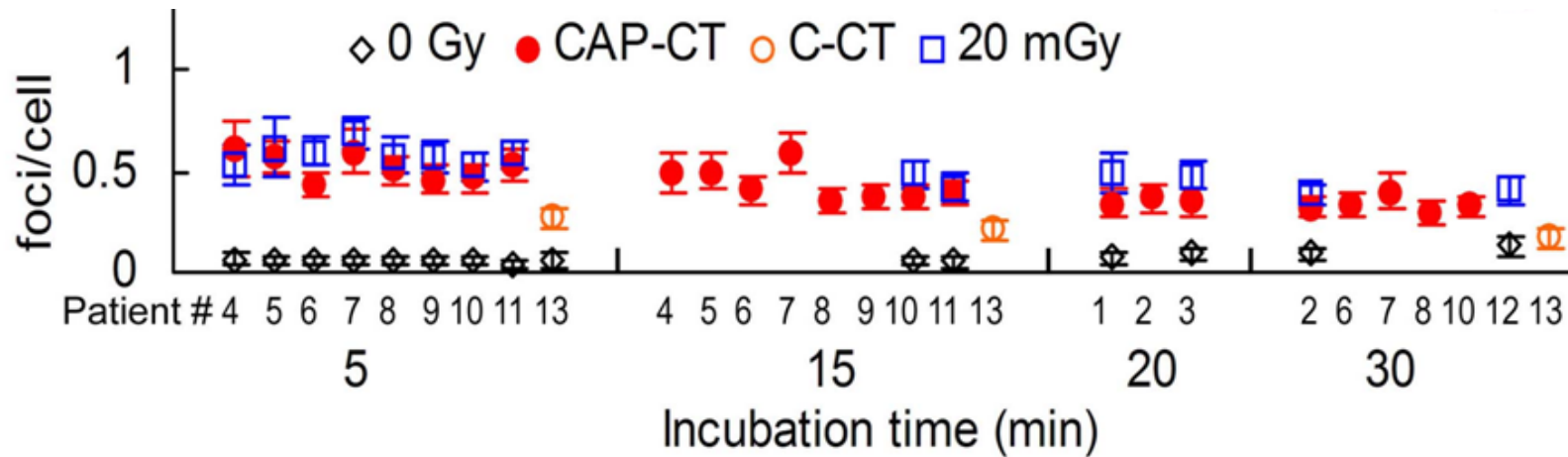


| Sample | radiation dose (Gy) | dose rate (mGy/min) | volume (mL) | current (mA) | exposure time (h) | temperature (°C) |
|---|---------------------|---------------------|-------------|--------------|-------------------|------------------|
| Calibration curve I | | | | | | |
| 1 | 0 | 0 | 0 | 0 | 8 | 37 |
| 2 | 0.25 | 0.3 | 200 | 1.2 | 0.5 | 37 |
| 3 | 0.5 | 0.3 | 200 | 1.2 | 1 | 37 |
| 4 | 1 | 0.3 | 200 | 1.2 | 2 | 37 |
| 5 | 2 | 0.3 | 200 | 1.2 | 4 | 37 |
| 6 | 4 | 0.3 | 200 | 1.2 | 8 | 37 |
| Calibration curve II | | | | | | |
| 1 | 0 | 0 | 0 | 0 | 4 | 37 |
| 2 | 0.125 | 0.5 | 120 | 1.2 | 4 | 37 |
| 3 | 0.25 | 1 | 120 | 1.2 | 4 | 37 |
| 4 | 0.5 | 2.1 | 180 | 1.2 | 4 | 37 |
| 5 | 1 | 4.2 | 180 | 1.2 | 4 | 37 |
| 6 | 2 | 8.3 | 200 | 1.2 | 8 | 37 |
| Actual samples | | | | | | |
| 1A | 2.4 | 40.0 | na | na | 1 | 37 |
| 1B | 0.56 | 1.5 | na | na | 1 | 37 |
| 2A | 1.6 | 17.8 | na | na | 2.5 | 37 |
| 5A | 0.25 | 2.8 | na | na | 2.5 | 37 |
| blot sample equivalent, incubated over 8h, 37°C | | | | | | |
| 1A-8h | 2.4 | 40 | 200 | 5.5 | 1+1* | 37 |
| 1B-8h | 0.2 | 3.3 | 175 | 1.2 | 1+1* | 37 |
| 2A-8h | 1.5 | 12.5 | 240 | 1.2 | 2+2* | 37 |
| 3A-8h | 0.2 | 1.7 | 150 | 1.2 | 2+2* | 37 |
| blot sample equivalent, incubated over 8h, alternating temperature 16-37°C | | | | | | |
| 1A-8-37°C | 2.4 | 40 | 200 | 5.5 | 1+1* | 37 |
| 1A-8-37°C | 2.4 | 40 | 200 | 5.5 | 1+1* | 37 |
| 1A-8-37°C | 2.4 | 40 | 200 | 5.5 | 1+1* | 37 |
| 1A-8-27°C | 2.4 | 40 | 200 | 5.5 | 1+1* | 27 |

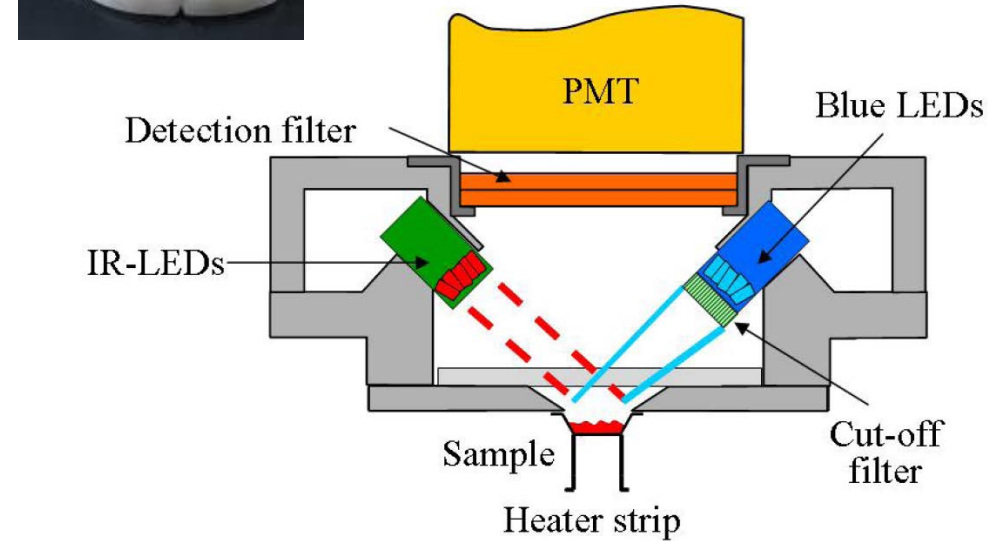


³-H2AX assay for biological dosimetry

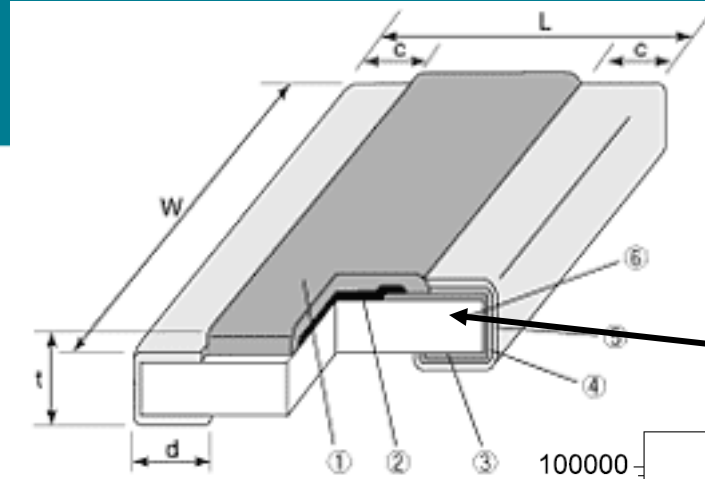
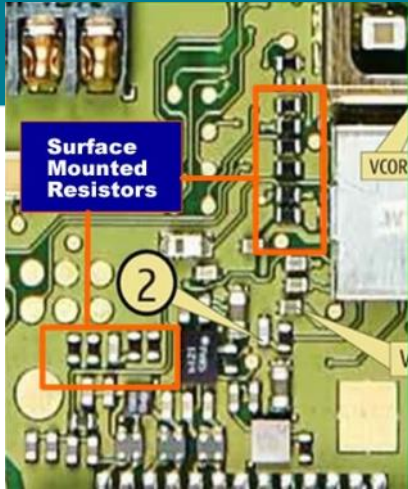
Blood samples taken after CT scans:



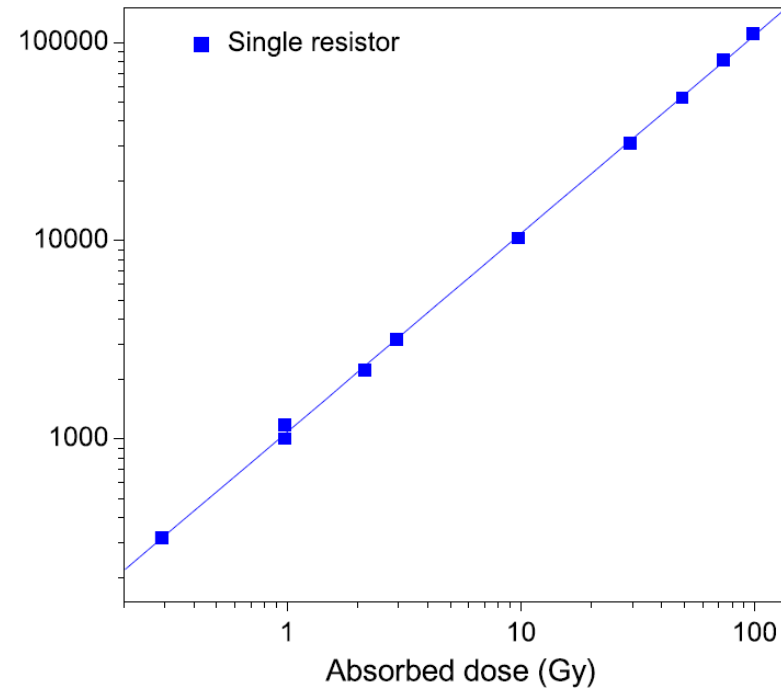
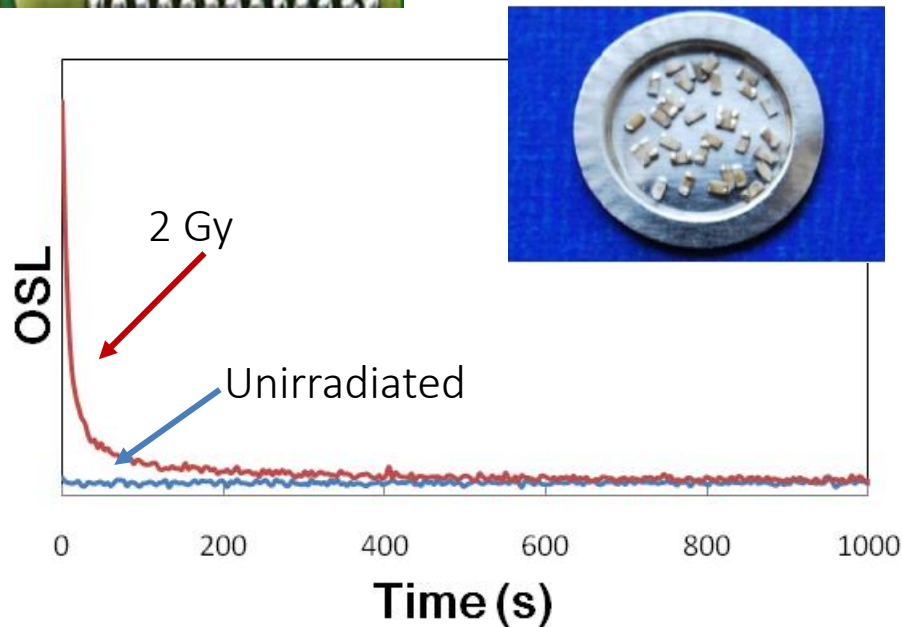
Thermo-/optically- stimulated luminescence



OSL - resistors



- (1) Protective coating
- (2) Resistive film
- (3) Inner electrode
- (4) Ni plating
- (5) Sn plating
- (6) **Ceramic substrate**



~10-20 mGy detectable but 50% fading in 10 days



Electron paramagnetic resonance



Fattibene & Callens (2010) Appl Radiat Isot 68:2033;
Trompier et al (2009) Ann Ist Super Sanita 45:287

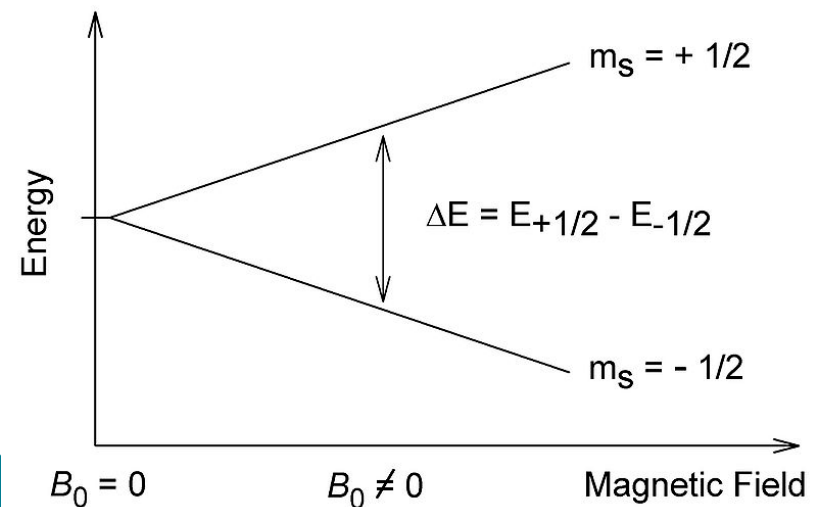
EPR dosimetry measures the concentration of radicals with unpaired electrons in a sample.

[carried out by measuring how much microwave energy is absorbed due to 'flipping' of electron spins in a magnetic field]

Radicals persist sufficiently long only in very few 'solid' biological materials:

- Calcified tissues (teeth, bones)
- Finger/toe nails
- Hair

...but also detectable in personal belongings, e.g. glass (watch, glasses, phone...)



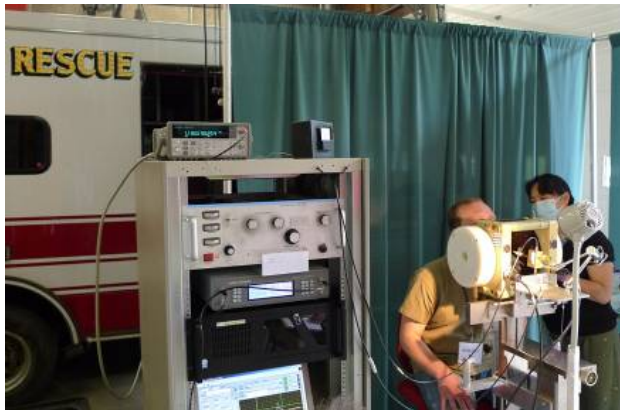
EPR *in vivo*: teeth

📖 Swartz et al (2006) Radiat Prot Dosimetry 120:163

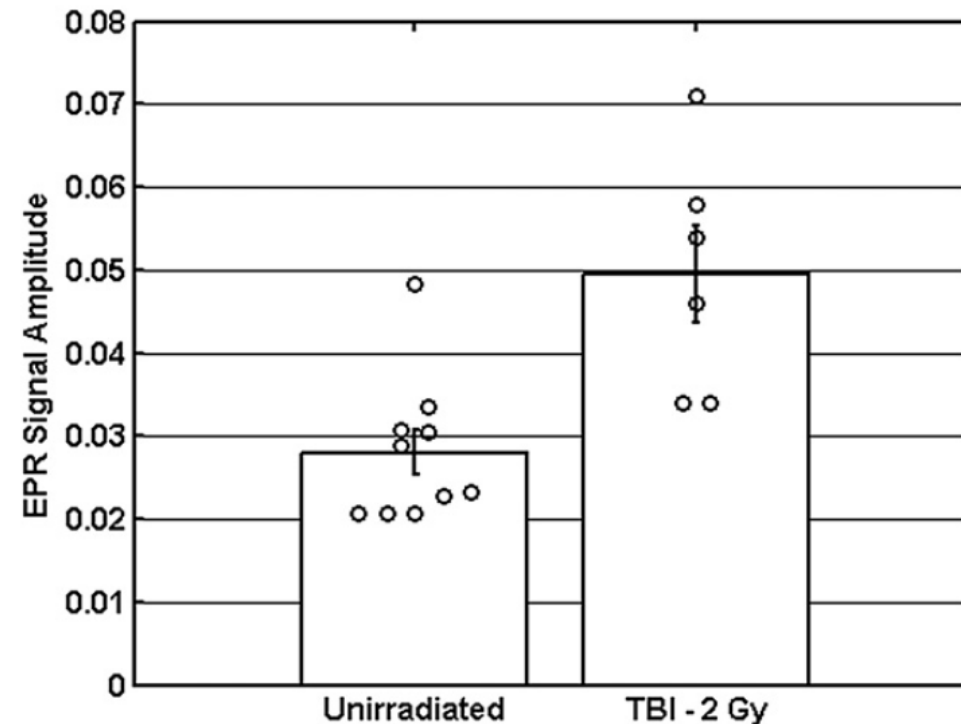
Williams et al (2011) Radiat Meas 46:772-7

Field deployable *in vivo* EPR tooth dosimeter being developed...

- 17 cm dipole magnet
- SE ~0.9 Gy
- Smaller versions planned



http://www.dartmouth.edu/~eprctr/research/tooth_dosimetry.shtml



Future work: Improve sensitivity further, develop smaller field-deployable systems suitable for triage

Mass casualty radiation accidents



1) Initial clinical triage

- vomiting, etc

2) Secondary triage

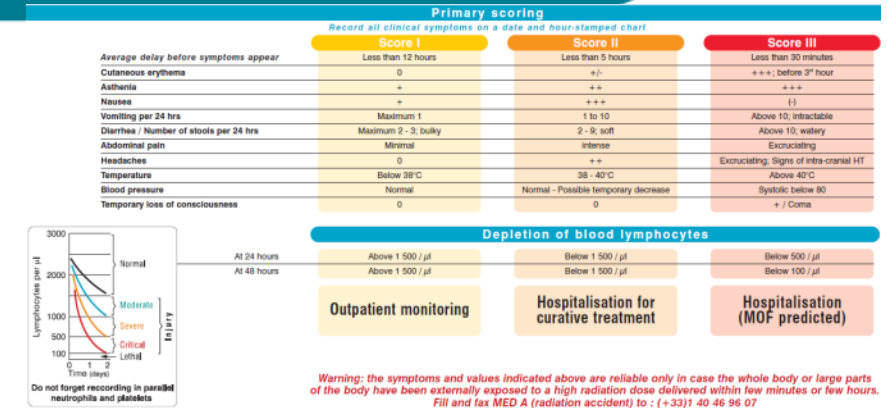
- retrospective dosimetry

Problem

Small very specialised laboratories with limited surge capacity

Solutions

- Score fewer cells at first (e.g. 50 instead of 500) (ISO 21243)
- 'Quickscan' (Flegal et al (2010) Health Phys 98:276)
- Automation
- New methods / combination of methods
- Assistance networks



Retrospective dosimetry assistance networks

Regional:

EU: RENE B (Kulka et al, Int J Radiat Biol. 2017 Jan;93(1):2-14); based on recent research projects
Latin America, Japan, Canada, ...

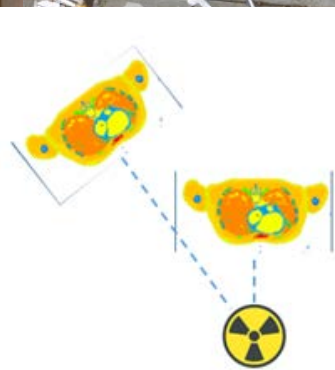


Worldwide:

International Atomic Energy Agency – Response Assistance Network (RANET)
World Health Organization – BioDoseNet



EURADOS WG 10



Venue
 IRSN, Bâtiment 03
 12 rue de la Justice
 Fontenay-Aux-Roses (close to Paris)
 France

• By metro Line 10 station Châtillon/Montrouge then Tramway T6 stop Division Leclerc
 • By train RER B (Charles de Gaulle/Robinson) station Fontenay-Aux-Roses then Bus 304 stop Division Leclerc

Further information & Registration
dosicourse@irsn.fr
<http://www.concert-h2020.eu/>

Organizing committee
 Sophie Ancelet (IRSN, France, Chair)
 Liz Ainsbury (PHE, UK)
 Christiane Wiele (Helmholtz-Zentrum für Umweltforschung, Germany)
 Augusto Giussani (BfS, Germany)

IRSN **EURADOS**
 Helmholtz Zentrum für Umweltforschung
 Deutscher Versuchsschutz **Public Englistar**

CONCERT
 This course is funded by the Europe Joint Programme for the Integrated Radiation Protection Research (CONCERT 2014 Education & Training)

Uncertainty in biological, physical and internal dosimetry following a single exposure
 April 15-19, 2019
 IRSN, Paris, France

SATELLITE EVENT

Training Course on "How to Measure and Analyze Luminescence Signals for Potential Applications in Radiation Dosimetry: Theory and Computational Procedures" (WG10)

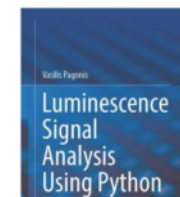
The training course takes place on Monday, June 12th, 08:00-12:30 and consists of two parts:

PART I: Steve McKeever, Emeritus Regents Professor of Physics, Oklahoma State University, USA

"Garbage In: Garbage Out". Understanding What You Measure is Critical

The discussion will cover understanding kinetics, what the standard equations mean and don't mean, how to collect the data, essential basic analytical tools, rudimentary and novel peak fitting approaches.

PART II: Vasilis Pagonis, Emeritus Professor of Physics, McDaniel College, USA and Associate Editor, Radiation Measurements



Luminescence signal analysis with open access software in Python and R

Practical examples will be demonstrated using software codes in Python and R for TL/OSL analysis, using actual experimental data files from dosimetric materials. Codes will be made available at the workshop in the form of Jupyter notebooks, which participants will be able to download freely from the web. Using their own laptops, participants can log into their Google Drive account and can run the codes immediately in

Joint EURADOS WG7 (Internal dosimetry) and WG10 (Retrospective dosimetry) workshop: "Biological Dosimetry techniques and EPR applied to accidental intakes of radionuclides"- open to all

9th October 2022, 13:30 – 18:00

Aim: To discuss the key open research questions and the potential to address these through collaborative research within and external to EURADOS.

Presentations will include:

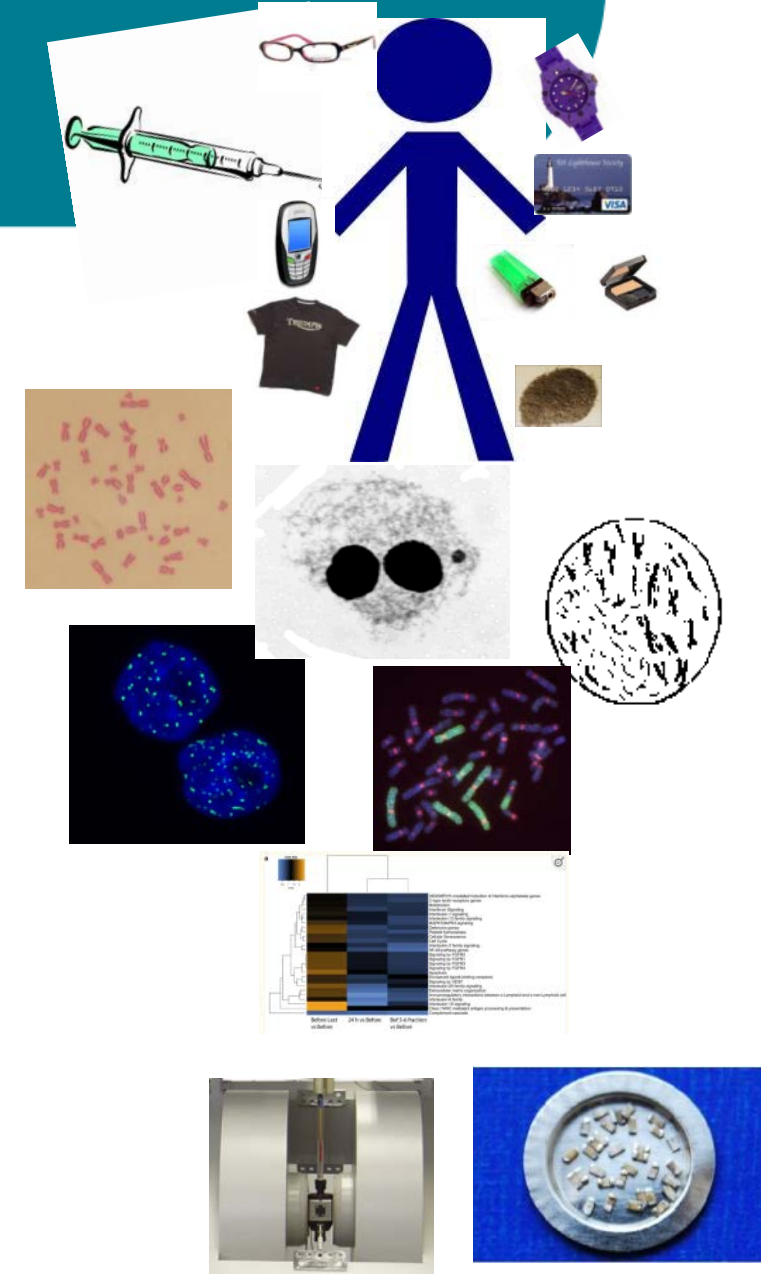
- Topic 1: **Biological and internal dosimetry for radiation medicine: current state and prospects for the future**, Satoshi Tashiro (Hiroshima University)
- Topic 2: **Establishment of calibration curves and intercomparison on calibration**, Liz Ainsbury (UKHSA)
- Topic 3: **Uncertainties in internal dosimetry/biodosimetry**, Augusto Giussani (Federal Office for Radiation Protection BfS) and Paco Barquinero (Universitat Autònoma de Barcelona)

The presentations will be followed by round table discussions on how to move forward with these topics.

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opic. A flyer

Summary

- Retrospective dosimetry quantifies radiation-induced changes to biological or certain physical 'fortuitous' materials carried by suspected exposed individuals
- Development started in the 1960s; today we have a robust set of tools and techniques to estimate dose and in some circumstances say something about circumstances of exposure
- Development continues!
- For large scale emergency response, networking in development and validation is absolutely key to ensuring we are able to respond with the state of the art for retrospective dose estimation
- EURADOS WG 10, RENEB and other networks remain essential – this has been and must remain an international effort



Thank you for listening!

All work/images not referenced are my own or presented on behalf of WG 10 – thank you to all colleagues

Questions or comments?
Interested to join WG10?

Do get in touch:

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